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(57) Abstract			
This invention describes genes encoding proteins which control resistance of plants to fungal pathogens. The invention also describes transgenic plants resistant to fungal pathogens and methods for making plants resistant to fungal pathogens. The invention further discloses a method to isolate additional genes coding for additional proteins controlling the resistance of plants to fungal pathogens.			

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GENES CONTROLLING DISEASES

The invention describes nucleotide sequences encoding proteins controlling resistance of plants to fungal diseases. The invention also relates to plants resistant to fungal diseases, and to methods of making plants resistant to fungal diseases.

Fungal diseases are responsible for yearly losses of about \$ 9.1 bn on agricultural crops in the US, and are caused by a large variety of biologically diverse pathogens. Different strategies have traditionally been used to control them. Resistance traits have been bred into agriculturally important varieties, thus providing various levels of resistance against either a narrow range of pathogen isolates or races, or against a broader range. However, this involves the long and labor intensive process of introducing desirable traits into commercial lines by genetic crosses and, due to the risk of pests evolving to overcome natural plant resistance, a constant effort to breed new resistance traits into commercial lines is required. Alternatively, fungal diseases have been controlled by the application of chemical fungicides. This strategy usually results in efficient control, but is also associated with the possible development of resistant pathogens and can be associated with a negative impact on the environment. Moreover, in certain crops, such as barley and wheat, the control of fungal pathogens by chemical fungicides is difficult or impractical.

Recent techniques have allowed a better understanding of the interactions between plants and their pathogens at the molecular level and mechanisms of resistance have been partially unraveled. While a large portion of this molecular characterization has been conducted in the model plant *Arabidopsis*, resistance mechanisms have also begun to be elucidated in economically important crops.

The powdery mildews are a major disease affecting most plant species and have been widely studied. They are characterized by spots or patches of a white to grayish growth on plant tissues, which correspond to the mycelium and cleistothecia of the fungus. Powdery mildews are caused by several species of fungi of the order Erysiphales. For example, *Erysiphe graminis* causes the powdery mildew of cereals and grasses. While powdery mildews are hard to control in most crops, barley lines resistant to most known isolates of the pathogen are available. It was shown that mutations at a single locus, the Mlo locus, are responsible for resistance phenotype. The mechanism of mlo resistance has been partially elucidated; it involves the formation of large cell wall appositions, called papillae, at the contact sites with the pathogen, which mainly contain callose, but also carbohydrates,

phenols and proteins. In mlo plants, cell wall appositions prevent the penetration of the pathogen, thus providing resistance.

Unfortunately, this powerful tool to control powdery mildews is restricted to barley. In the view of the problems caused by fungal diseases in agriculture, in particular by powdery mildews, there remains an unfulfilled need for new and effective strategies to control these types of pathogens in other crops, which are economically attractive to the farmers and environmentally acceptable.

The present invention addresses the need for novel disease control strategies in plants via the application of genetic engineering techniques. In particular, this invention relates to control strategies against powdery mildew, preferably in economically important crops. The present invention relates to isolated DNA molecules encoding Mlo proteins, wherein such Mlo proteins confer resistance of plants to fungal pathogens. In particular, the invention relates to Mlo proteins containing conserved amino acid sequences that the inventors of the present invention are first to discover, and to the isolated DNA molecules encoding such Mlo proteins. The present invention is also drawn to vectors for expression of the DNA molecules of the present invention in plants. The present invention further relates to transgenic plants comprising any one of the DNA molecules of the present invention. The present invention also describes agricultural products with improved phytosanitary properties comprising transgenic plants resistant to fungal pathogens by expression of any of the DNA molecules of the present invention. The present invention also further relates to methods of making plants resistant to fungal diseases by altering the expression in transgenic plants of proteins encoded by the endogenous copies of the genes corresponding to any one of the DNA molecules of the present invention or altering the activity or the stability of proteins encoded by the endogenous copies of the genes corresponding to any one of the DNA molecules of the present invention. Such transgenic plants are desirably resistant to pathogens that infect living epidermal plant cells, more desirably to fungi from the order Erysiphales, also known as powdery mildews, preferably from the genus *Erysiphe*, the causing agent of powdery mildew, more preferably the plants are resistant to *Erysiphe graminis*. The present invention further describes a method to isolate DNA molecules encoding proteins having the same or a similar function as the DNA molecules of the present invention and encoding the conserved amino acid sequences set forth in the present invention.

The present invention thus provides new and effective strategies to control fungal diseases in economically important crops, potentially reducing amounts of chemicals applied to crops and reducing the risk of appearance of pathogens resistant to control agents.

The invention thus provides:

A DNA molecule encoding an Mlo protein conferring upon a plant resistance to fungal pathogens, wherein said protein comprises at least one amino acid sequence identical or substantially similar to an amino acid sequence set forth in SEQ ID No:1 or SEQ ID No:2, wherein said DNA molecule is preferably a cDNA molecule. In a preferred embodiment, the DNA molecule is preferably not derived from barley and is derived from a plant which is either a dicot or from a group of plants consisting of wheat, corn, rice, oats, rye, sorghum, sugarcane, millet, milo, and the palm family. In a preferred embodiment, the DNA molecule of the present invention is identical or substantially similar to any one of the nucleotide sequences set forth in SEQ ID No: 3, SEQ ID No: 5 or SEQ ID No:7, or encodes an Mlo protein identical or substantially similar to an Mlo protein set forth in SEQ ID No: 4, SEQ ID No: 6 or SEQ ID No: 8. In a more preferred embodiment, the DNA molecule comprising the nucleotide sequences set forth in SEQ ID No: 3, SEQ ID No: 5 or SEQ ID No: 7 is derived from wheat. In another preferred embodiment, the DNA molecule of the present invention is identical or substantially similar to any one of the nucleotide sequences set forth in SEQ ID No: 9, SEQ ID No: 11, SEQ ID No: 13, SEQ ID No:15 or SEQ ID No: 17, or encodes an Mlo protein identical or substantially similar to an Mlo protein encoded by any one of the nucleotide sequences set forth in SEQ ID No: 10, SEQ ID No: 12, SEQ ID No: 14, SEQ ID No:16 or SEQ ID No:18. In a more preferred embodiment, the DNA molecule comprising the nucleotide sequences set forth in SEQ ID No: 9, SEQ ID No: 11, SEQ ID No: 13, SEQ ID No:15 or SEQ ID No: 17 is derived from *Arabidopsis thaliana*. In another preferred embodiment, said DNA molecules mentioned hereinbefore are modified such that the activity of the endogenous protein is lost. In one particular embodiment of the present invention, said DNA modification results in one, all or a combination of the following changes in the amino acid sequence of the corresponding protein

- Trp (163) to Arg
- frameshift after Pro (396)
- frameshift after Trp (160)
- Met (1) to Ile

- Gly (227) to Asp
- Met (1) to Val
- Arg (11) to Trp
- missing Phe (183), Thr (184)
- Val (31) to Glu
- Ser (32) to Phe
- Leu (271) to His.

In a further preferred embodiment, the fungal pathogens desirably infect living epidermal cells, more desirably the fungal pathogens are from the order Erysiphales, also known as powdery mildews, preferably from the genus *Erysiphe* and more preferably the fungal pathogen is *Erysiphe graminis*.

In a further embodiment, the isolated DNA molecule is antisense to an isolated molecule as described above, e.g. antisense to a DNA molecule, e.g. a cDNA molecule, encoding an Mlo protein comprising at least one amino acid sequence identical or substantially similar to an amino acid sequences set forth in SEQ ID No:1 or SEQ ID No:2, especially antisense to a DNA molecule identical or substantially similar to a DNA molecule set forth in SEQ ID Nos:3, 5, 7, 9, 11, 13, 15 or 17 and encoding an Mlo protein identical or substantially similar to an Mlo protein set forth in SEQ ID Nos:4, 6, 8, 10, 12, 14, 16 or 18.

The invention further provides:

A protein comprising at least one amino acid sequence identical or substantially similar to an amino acid sequence set forth in SEQ ID No: 1 or SEQ ID No:2, wherein said protein is an Mlo protein and confers upon a plant resistance to fungal pathogens. The protein is preferably not derived from barley and is derived from a plant which is either a dicot or from a group of plants consisting of wheat, corn, rice, oats, rye, sorghum, sugarcane, millet, milo, and the palm family. In a preferred embodiment, the protein of the present invention is encoded by a nucleotide sequence identical or substantially similar to any one of the nucleotide sequences set forth in SEQ ID No: 3, SEQ ID No: 5 or SEQ ID No:7 or is identical or substantially similar to any one of the Mlo proteins set forth in SEQ ID No: 4, SEQ ID No: 6 or SEQ ID No: 8. In a more preferred embodiment, the protein is derived from wheat. In another preferred embodiment, the protein of the present invention is encoded by a nucleotide sequence identical or substantially similar to any one of the nucleotide sequences set forth in SEQ ID No: 9, SEQ ID No: 11, SEQ ID No: 13, SEQ ID No:15 or

SEQ ID No: 17 or is identical or substantially similar to any one of the Mlo proteins set forth in SEQ ID No: 10, SEQ ID No: 12, SEQ ID No: 14, SEQ ID No: 16 or SEQ ID No: 18. In a more preferred embodiment, the protein is derived from *A. thaliana*. In another preferred embodiment, the fungal pathogens desirably infect living epidermal cells, more desirably the fungal pathogens are from the order Erysiphales, also known as powdery mildews, preferably from the genus *Erysiphe* and more preferably the fungal pathogen is *Erysiphe graminis*. In a further embodiment, the present invention also encompasses mutated forms or truncated forms of proteins encoded by any of the DNA molecules described above.

The invention further provides:

An expression cassette comprising any one of the DNA molecules described above, e.g. a cDNA as described above, wherein the DNA molecule is operably linked to a promoter and to termination signals capable of expressing the DNA molecule in plants. In a preferred embodiment, the expression cassette is heterologous. In a further preferred embodiment, the promoter and the termination signals are eukaryotic. In a further preferred embodiment, the promoter and termination signals are heterologous with respect to the coding region.

The invention further provides:

A vector comprising any one of the expression cassettes described above. In a preferred embodiment, the vector is used for transformation of the expression cassette in plants. In another preferred embodiment, the vector of the present invention is used for amplification of any one of the DNA molecules described above.

The invention further provides:

A cell comprising an expression cassette or parts of it comprising an isolated DNA molecule of the present invention, wherein said DNA molecule in said expression cassette is expressible in said cell. In a preferred embodiment, the DNA molecule is not derived from barley. In another preferred embodiment, the cell is a plant cell. In a further preferred embodiment, the expression cassette is stably integrated in the genome of the cell or is included in a self-replicating vector and remains in the cell as an extrachromosomal molecule.

The invention further provides:

A plant comprising an expression cassette or parts of it comprising an isolated DNA molecule of the present invention. In a preferred embodiment, the DNA molecule is not derived from barley. In another preferred embodiment, the DNA molecule comprised in the expression cassette is expressible in the plant. In another preferred embodiment, the DNA molecule is stably integrated in the plant genome or is included in a self-replicating vector and remains in the cell as an extrachromosomal molecule. In another preferred embodiment, the plant is resistant to fungal pathogens, desirably fungal pathogens which infect living epidermal cells, more desirably the fungal pathogens are from the order Erysiphales, also known as powdery mildews, preferably from the genus *Erysiphe* and more preferably the fungal pathogen is *Erysiphe graminis*.

The inventions also relates to the seed for such a plant, which seed is optionally treated (e.g. primed or coated) and/or packaged, e.g. placed in a bag with instructions for use.

The invention further provides:

Agricultural products comprising a plant comprising an isolated DNA molecule of the present invention. In a preferred embodiment, the agricultural product is used as e.g. feed, food, or silage and does not contain mycotoxins produced by fungal pathogens, such as e.g. aflatoxins. Therefore, the agricultural product has improved phytosanitary properties.

The invention further provides:

A method for making a plant resistant to a fungal pathogen comprising the step of:

- a) expressing in a plant an RNA transcript encoded by any one of the DNA molecules described above in "sense" orientation; or
- b) expressing in a plant an RNA transcript encoded by any one of the DNA molecules described above in "anti-sense" orientation; or
- c) expressing in a plant a ribozyme capable of specifically cleaving a messenger RNA transcript encoded by an endogenous gene corresponding to any one of the DNA molecules described above; or
- d) expressing in a plant an aptamer specifically directed to an endogenous protein encoded by a gene corresponding to any one of the DNA molecules described above; or
- e) expressing in a plant a mutated or a truncated form of any one of the DNA molecules described above, so that it can act as a dominant negative mutant; or
- f) modifying by homologous recombination in a plant at least one chromosomal copy of the gene corresponding to any one of the DNA molecules described above; or

g) modifying by homologous recombination in a plant at least one chromosomal copy of the regulatory elements of a gene corresponding to any one of the DNA molecules described above.

The invention further provides:

A plant obtained by any one of the method described immediately above including the seed for such a plant, which seed is optionally treated (e.g., primed or coated) and/or packaged, e.g. placed in a bag with instructions for use. In another preferred embodiment, the plant obtained is resistant to fungal pathogens, desirably fungal pathogens which infect living epidermal cells, more desirably the fungal pathogens are from the order Erysiphales, also known as powdery mildews, preferably from the genus *Erysiphe* and more preferably the fungal pathogen is *Erysiphe graminis*.

The invention further provides:

An agricultural product with improved phytosanitary properties obtained by any one of the methods described immediately above.

The invention further provides:

A method for isolating DNA molecules encoding Mlo proteins comprising the steps of:

- a) mixing a degenerated oligonucleotide encoding at least six amino acids of SEQ ID No:1 and a degenerated oligonucleotide complementary to a sequence encoding at least six amino acids of SEQ ID No:2, with DNA extracted from a plant under conditions allowing hybridization of said degenerated oligonucleotides to said DNA; and
- b) amplifying a DNA fragment of said plant DNA, wherein said DNA fragment comprises a at its left and right ends nucleotide sequences that can anneal to said degenerated oligonucleotides in step a); and
- c) obtaining a full-length cDNA clone comprising the DNA fragment of step b).

The invention further provides:

A method for producing mutated copies of the nucleotide sequences of the present invention by "in-vitro recombination" or "DNA shuffling". The mutated copies of the nucleotide sequences of the present invention are used to confer improved resistance to fungal pathogens. In a preferred embodiment, the mutant copies of the nucleotide

sequences of the present invention are used to confer resistance to a broader range of pathogens. One such method is described below:

A method for mutagenizing a DNA molecule according to the present invention, wherein said DNA molecule has been cleaved into double-stranded-random fragments of a desired size, and comprising the steps of:

- a) adding to the resultant population of double-stranded random fragments one or more single or double-stranded oligonucleotides, wherein said oligonucleotides comprise an area of identity and an area of heterology to the double-stranded template polynucleotide;
- b) denaturing the resultant mixture of double-stranded random fragments and oligonucleotides into single-stranded fragments;
- c) incubating the resultant population of single-stranded fragments with a polymerase under conditions which result in the annealing of said single-stranded fragments at said areas of identity to form pairs of annealed fragments, said areas of identity being sufficient for one member of a pair to prime replication of the other, thereby forming a mutagenized double-stranded polynucleotide; and
- d) repeating the second and third steps for at least two further cycles, wherein the resultant mixture in the second step of a further cycle includes the mutagenized double-stranded polynucleotide from the third step of the previous cycle, and the further cycle forms a further mutagenized double-stranded polynucleotide.

DEFINITIONS

An "isolated DNA molecule" is a nucleotide sequence that, by the hand of man, exists apart from its native environment and is therefore not a product of nature. An isolated nucleotide sequence may exist in a purified form or may exist in a non-native environment such as, for example, a transgenic host cell.

A "protein" as defined herein is the entire protein encoded by the corresponding nucleotide sequence, or is a portion of the protein encoded by the corresponding portion of the nucleotide sequence.

An "isolated protein" is a protein that is encoded by an isolated nucleotide sequence and is therefore not a product of nature. An isolated protein may exist in a purified form or may exist in a non-native environment, such as a transgenic host cell, wherein the protein would

not normally expressed or would be expressed in a different form or different amount in an isogenic non-transgenic host cell.

A plant "resistant to a fungal pathogen" has no or lesser symptoms of a fungal infection caused by the fungal , by inhibiting or limiting the ability of the fungal pathogen to grow on the plant. As a consequence, the plant grows better, has higher yields and produces more seeds.

"A protein conferring resistance upon a plant to a fungal pathogen" means that the protein is involved in the regulation of plant genetic pathways responsible for resistance of the plant to the fungal pathogen. The protein may be a positive regulator in that it enhances resistance of the plant to the fungal pathogen, or the protein may be a negative regulator in that it represses resistance of the plant to the fungal pathogen. A particular example of a protein conferring resistance upon a plant to a fungal pathogen is an Mlo protein.

An "Mlo protein" means herein a member of a family of proteins (the Mlo family) having a substantially similar function in a disease resistance pathway and sharing some structural homology. The structural homology can be e.g. that the members of the family share at least one conserved region.

In its broadest sense, the term "substantially similar", when used herein with respect to a nucleotide sequence, means a nucleotide sequence corresponding to a reference nucleotide sequence, wherein the corresponding sequence encodes a polypeptide having substantially the same structure and function as the polypeptide encoded by the reference nucleotide sequence, e.g. where only changes in amino acids not affecting the polypeptide function occur. Desirably the substantially similar nucleotide sequence encodes the polypeptide encoded by the reference nucleotide sequence. The percentage of identity between the substantially similar nucleotide sequence and the reference nucleotide sequence desirably is at least 80%, more desirably 85%, preferably at least 90%, more preferably at least 95%, still more preferably at least 99%.

The term "substantially similar", when used herein with respect to a protein, means a protein corresponding to a reference protein, wherein the protein has substantially the same structure and function as the reference protein, e.g. where only changes in amino acids not

affecting the polypeptide function occur. When used for a protein or an amino acid sequence the percentage of identity between the substantially similar and the reference protein or amino acid sequence desirably is at least 80%, more desirably 85%, preferably at least 90%, more preferably at least 95%, still more preferably at least 99%.

The percentage of sequence identity is determined using computer programs that are based on dynamic programming algorithms. Computer programs that are preferred within the scope of the present invention include the BLAST (Basic Local Alignment Search Tool) search programs designed to explore all of the available sequence databases regardless of whether the query is protein or DNA. Version BLAST 2.0 (Gapped BLAST) of this search tool has been made publicly available on the Internet (currently <http://www.ncbi.nlm.nih.gov/BLAST/>). It uses a heuristic algorithm which seeks local as opposed to global alignments and is therefore able to detect relationships among sequences which share only isolated regions. The scores assigned in a BLAST search have a well-defined statistical interpretation. Said programs are preferably run with optional parameters set to the default values.

The term "gene" refers to a coding sequence and associated regulatory sequences wherein the coding sequence is transcribed into RNA such as mRNA, rRNA, tRNA, snRNA, sense RNA or antisense RNA. Examples of regulatory sequences are promoter sequences, 5' and 3' untranslated sequences and termination sequences. Further elements that may be present are, for example, introns.

"Expression" refers to the transcription and/or translation of an endogenous gene or a transgene in plants. In the case of antisense constructs, for example, expression may refer to the transcription of the antisense DNA only.

"Expression cassette" as used herein means a DNA sequence capable of directing expression of a particular nucleotide sequence in an appropriate host cell, comprising a promoter operably linked to the nucleotide sequence of interest which is operably linked to termination signals. It also typically comprises sequences required for proper translation of the nucleotide sequence. The coding region usually codes for a protein of interest but may also code for a functional RNA of interest, for example antisense RNA or a nontranslated RNA that, in the sense or antisense direction, inhibits expression of a particular gene, e.g., antisense RNA. The expression cassette comprising the nucleotide sequence of interest

may be chimeric, meaning that at least one of its components is heterologous with respect to at least one of its other components. The expression cassette may also be one which is naturally occurring but has been obtained in a recombinant form useful for heterologous expression. Typically, however, the expression cassette is heterologous with respect to the host, i.e., the particular DNA sequence of the expression cassette does not occur naturally in the host cell and must have been introduced into the host cell or an ancestor of the host cell by a transformation event. The expression of the nucleotide sequence in the expression cassette may be under the control of a constitutive promoter or of an inducible promoter which initiates transcription only when the host cell is exposed to some particular external stimulus. In the case of a multicellular organism, such as a plant, the promoter can also be specific to a particular tissue or organ or stage of development.

"Heterologous" as used herein means "of different natural or of synthetic origin" or represent a non-natural state. For example, if a host cell is transformed with a nucleic acid sequence derived from another organism, particularly from another species, that gene is heterologous with respect to that host cell and also with respect to descendants of the host cell which carry that gene. The transforming nucleic acid may comprise a heterologous promoter, heterologous coding sequence, or heterologous termination sequence. Alternatively, the transforming nucleic acid may be completely heterologous or may comprise any possible combination of heterologous and endogenous nucleic acid sequences. Similarly, heterologous refers to a nucleotide sequence derived from and inserted into the same natural, original cell type, but which is present in a non-natural state, e.g. a different copy number, or under the control of different regulatory elements.

The term "promoter" refers to a DNA sequence that initiates transcription of an associated DNA sequence. The promoter region may also include elements that act as regulators of gene expression such as activators, enhancers, and/or repressors.

"Synthetic nucleotide sequence" as used herein means a nucleotide sequence comprising structural characters that are not present in the natural sequence. For example, an artificial sequence that resembles more closely the G+C content and the normal codon distribution of dicot and/or monocot genes is said to be synthetic.

A regulatory DNA sequence is said to be "operably linked to" or "associated with" a DNA sequence that codes for an RNA or a protein if the two sequences are situated such that the regulatory DNA sequence affects expression of the coding DNA sequence.

"Regulatory elements" refer to sequences involved in conferring the expression of a nucleotide sequence. Regulatory elements comprise a promoter operably linked to the nucleotide sequence of interest and termination signals. They also typically encompass sequences required for proper translation of the nucleotide sequence.

A "plant" refers to any plant or part of a plant and particularly to seed plants at any stage of development. Therein are also included cuttings, cell or tissue cultures and seeds. As used in conjunction with the present invention, the term "plant tissue" includes, but is not limited to, whole plants, plant organs, plant seeds, protoplasts, callus, cell cultures, and any groups of plant cells organized into structural and/or functional units.

A "plant cell" refers to the structural and physiological unit of the plant, comprising a protoplast and a cell wall. The plant cell may be in form of an isolated single cell or a cultured cell, or as a part of higher organized unit such as, for example, a plant tissue, or a plant organ.

"Transformation" as used herein means introduction of a nucleic acid into a cell. In particular, the stable integration of a DNA molecule into the genome of an organism of interest.

A "selectable marker" is conferred by a gene whose expression in a plant cell gives the cell a selective advantage. The selective advantage possessed by the cells transformed with the selectable marker gene may be due to their ability to grow in the presence of a negative selective agent, such as an antibiotic or a herbicide, compared to the growth of non-transformed cells. The selective advantage possessed by the transformed cells, compared to non-transformed cells, may also be due to their enhanced or novel capacity to utilize an added compound as a nutrient, growth factor or energy source. Selectable marker gene also refers to a gene or a combination of genes whose expression in a plant cell gives the cell both, a negative and a positive selective advantage.

A "screenable marker" is conferred by a gene whose expression does not give a selective advantage to a transformed cell, but whose expression makes the transformed cell phenotypically distinct from untransformed cells.

BRIEF DESCRIPTION OF THE SEQUENCES IN THE SEQUENCE LISTING

SEQ ID No:1	conserved amino acid sequence 1
SEQ ID No:2	conserved amino acid sequence 2
SEQ ID No:3	nucleotide sequence of wheat Mlo protein TrMlo1
SEQ ID No:4	protein sequence of TrMlo1
SEQ ID No:5	nucleotide sequence of wheat Mlo protein TrMlo2
SEQ ID No:6	protein sequence of TrMlo2
SEQ ID No:7	nucleotide sequence of wheat Mlo protein TrMlo3
SEQ ID No:8	protein sequence of TrMlo3
SEQ ID No:9	nucleotide sequence of Arabidopsis Mlo protein CIB10259
SEQ ID No:10	protein sequence of CIB10259
SEQ ID No:11	nucleotide sequence of Arabidopsis Mlo protein CIB10295
SEQ ID No:12	protein sequence of CIB10295
SEQ ID No:13	nucleotide sequence of Arabidopsis Mlo protein CIB10296
SEQ ID No:14	protein sequence of CIB10296
SEQ ID No:15	nucleotide sequence of Arabidopsis Mlo protein F19850
SEQ ID No:16	protein sequence of F19850
SEQ ID No:17	nucleotide sequence of Arabidopsis Mlo protein U95973
SEQ ID No:18	protein sequence of U95973
SEQ ID No:19	oligonucleotide MLO-1
SEQ ID No:20	oligonucleotide MLO-3
SEQ ID No:21	oligonucleotide MLO-5
SEQ ID No:22	oligonucleotide MLO-7
SEQ ID No:23	oligonucleotide MLO-10
SEQ ID No:24	oligonucleotide MLO-15
SEQ ID No:25	oligonucleotide MLO-26
SEQ ID No:26	oligonucleotide MLO-GSP1

SEQ ID No:27 oligonucleotide MLO-GSP2
SEQ ID No:28 oligonucleotide ST27
SEQ ID No:29 oligonucleotide N37544-1
SEQ ID No:30 oligonucleotide N37544-2
SEQ ID No:31 oligonucleotide T22146-1
SEQ ID No:32 oligonucleotide T22146-2
SEQ ID No:33 oligonucleotide H76041-1
SEQ ID No:34 oligonucleotide H76041-2
SEQ ID No:35 oligonucleotide SAS-1
SEQ ID No:36 oligonucleotide SAS-2
SEQ ID No:37 oligonucleotide SAS-3
SEQ ID No:38 oligonucleotide SAS-4
SEQ ID No:39 oligonucleotide SAS-5
SEQ ID No:40 oligonucleotide SAS-6
SEQ ID No:41 oligonucleotide SAS-7
SEQ ID No:42 oligonucleotide SAS-8

Deposits

Deposited material	Accession number	Date of deposit
pCIB 10259	NRRL B-21945	3/10/98
pCIB 10295	NRRL B-21946	3/10/98
pCIB 10296	NRRL B-21947	3/10/98
TrMlo1	NRRL B-21948	3/10/98
TrMlo1-5	NRRL B-21949	3/10/98
TrMlo2	NRRL B-21950	3/10/98
TrMlo2-5	NRRL B-21951	3/10/98
TrMlo3	NRRL B-21952	3/10/98
TrMlo3-5	NRRL B-21953	3/10/98

All deposits were made with the Northern Regional Research Center, 1815 Northern University Street, Peoria, Illinois 61604, USA.

The present invention relates to DNA molecules encoding Mlo proteins, which confer upon a plant resistance to fungal pathogens. The inventors of the present invention are first to identify conserved amino acid sequences among Mlo proteins. The conserved amino acid sequences of the present invention are conserved between three Mlo proteins derived from wheat and three Mlo proteins derived from *A. thaliana*. These amino acid sequences are also conserved in two predicted Arabidopsis Mlo proteins. The first conserved amino acid sequence, which is set forth in SEQ ID No:1, comprises 13 amino acids. The fourth amino acid in SEQ ID No:1 is either L, V or I, its fifth amino acid is either V or L and its seventh amino acid is either F or L. The thirteenth amino acid in SEQ ID No:1 is not I and is preferably either T, S or A. The second conserved amino acid sequence, which is set forth in SEQ ID No:2, comprises 14 amino acids. The first amino acid in SEQ ID No:2 is not M and is preferably either I, V, S or G. Its third amino acid is either F, L or V, its sixth amino acid is either Y or N, its seventh amino acid is either A or V, its eighth amino acid is either L or I, its tenth amino acid is either T or S. The invention encompasses isolated Mlo proteins comprising at least one of the conserved amino acid sequences described above and isolated DNA molecules which encode such Mlo proteins. The invention also encompasses isolated Mlo proteins which comprise both conserved sequences set forth in SEQ ID No: 1 and SEQ ID No:2. In a preferred embodiment, the isolated DNA molecules encoding the Mlo proteins of the present invention are cDNA molecules.

In a further embodiment, the DNA molecules encoding Mlo proteins comprising at least one of the conserved amino acid sequences are not derived from barley. In another embodiment, such DNA molecules are derived from a dicot or from wheat, corn, rice, oats, rye, sorghum, sugarcane, millet, milo, or the palm family. In preferred embodiment, a DNA molecule of the present invention is identical or substantially similar to the DNA molecules set forth in SEQ ID No:3, 5 or 7 and in SEQ ID No:9, 11, 13, 15 or 17 or encode an Mlo protein, which is identical or substantially similar to any one of the Mlo proteins set forth in SEQ ID No:4, 6 or 8 or in SEQ ID No:10, 12, 14, 16, 18. The DNA molecules set forth in SEQ ID No:3, 5 or 7 are derived from wheat and encode the Mlo proteins set forth in SEQ ID No:4, 6 or 8, respectively. The isolation of such DNA molecules is further illustrated in example 1. The DNA molecules set forth in SEQ ID No:9, 11, 13, 15 or 17 are derived from Arabidopsis and encode the Mlo proteins set forth in SEQ ID No:10, 12, 14, 16 or 18, respectively. The isolation of such DNA molecules is further illustrated in example 2.

The DNA molecule of SEQ ID No:3 encoding a wheat Mlo protein called TrMlo1 is deposited as strains TrMlo1 and TrMlo1-5 with accession numbers NRRL B-21948 and NRRL B-

21949, respectively. The DNA molecule of SEQ ID No:5 encoding a wheat Mlo protein called TrMlo2 is deposited as strains TrMlo2 and TrMlo2-5 with accession numbers NRRL B-21950 and NRRL B-21951, respectively. The DNA molecule of SEQ ID No:7 encoding a wheat Mlo protein called TrMlo3 is deposited as strains TrMlo3 and TrMlo3-5 with accession numbers NRRL B-21952 and NRRL B-21953, respectively. TrMlo1 and TrMlo3 comprise the full-length cDNAs of the corresponding Mlo genes and also comprise some of the corresponding 5' and 3' untranslated regions. TrMlo2 is the longest cDNA clone of the corresponding gene that was recovered. It comprises the entire coding region with exception of the first methionin (start codon) as deduced from a comparison with TrMlo1 and TrMlo3. TrMlo2 also comprises some of the 3' untranslated region of the corresponding gene.

The DNA molecule of SEQ ID No:9 encoding an Arabidopsis Mlo protein called CIB10259 is deposited as strain pCIB10259 with accession number NRRL B-21945. The DNA molecule of SEQ ID No:11 encoding an Arabidopsis Mlo protein called CIB10295 is deposited as strain pCIB10295 with accession number NRRL B-21946. The DNA molecule of SEQ ID No:13 encoding an Arabidopsis Mlo protein called CIB10296 is deposited as strain pCIB1029 with accession number NRRL B-21947. CIB10259, CIB10295 and CIB10296 comprise the full-length cDNAs of the corresponding Mlo genes and also comprise and also comprise some of the corresponding 5' and 3' untranslated regions. The nucleotide sequences encoding the Arabidopsis Mlo protein family members F19850 and U95973 are obtained from the Genbank. However, for both clones, a predicted amino acid sequence is determined and is found not to match the amino acid sequence predicted in the Genbank entry. The Mlo proteins determined by the inventors of the present invention are therefore novel and not obvious. Both newly predicted proteins contain the conserved amino acid sequences set forth in SEQ ID No:1 and 2 and are therefore encompassed by the present invention as well as the isolated cDNAs encoding them.

An Mlo protein encoded by a DNA molecule of the present invention confer upon a plant resistance to fungal pathogens, desirably fungal pathogens which infect living epidermal plant cells, more desirably fungal pathogens from the order Erysiphales, also known as powdery mildews (Agrios G. (1988) Plant Pathology, Third Edition, Academic Press Inc., in particular p 271). Preferably, an Mlo protein encoded by a DNA molecule of the present invention confer upon a plant resistance from the genus *Erysiphe*, more preferably the fungal pathogen is *Erysiphe graminis*.

The present invention also encompasses recombinant vectors comprising any one of the DNA molecules of this invention. In these vectors, such DNA molecules are preferably comprised in an expression cassette comprising regulatory elements for expression of the DNA molecules in a host cell capable of expressing such DNA molecules. Such regulatory elements are usually a promoter and termination signals and preferably also include elements allowing efficient translation of a protein encoded by a DNA molecule of the present invention. In a preferred embodiment, an expression cassette is heterologous. Such vectors are used for transformation of the expression cassette comprising any one of the DNA molecules of this invention into a host cells. In a preferred embodiment, the expression cassette is stably integrated into the DNA of such host cell. In another preferred embodiment, the expression cassette is comprised in a vector, which is capable of replication in a host cell and remains in the host cell as an extrachromosomal molecule. In a further preferred embodiment, such extrachromosomal replicating molecule is used to amplify the DNA molecules of this invention in a host cell. In a preferred embodiment, such host cell is a microorganism, such as a bacteria, in particular *E. coli*. In another preferred embodiment, a host cell is an eukaryotic cell, such as e.g. a yeast cell, an insect cell, or a plant cell.

In a further embodiment, a DNA molecule of the present invention is modified by incorporation of random mutations in a technique known as *in-vitro* recombination or DNA shuffling. This technique is described in Stemmer et al., Nature 370: 389-391 (1994) and US Patent 5,605,793 incorporated herein by reference. Millions of mutant copies of the nucleotide sequences are produced based on the original nucleotide sequence described herein and variants with improved properties, such as increased resistance for fungal pathogens or resistance against a wider range of pathogens are recovered. The method encompasses forming a mutagenized double-stranded polynucleotide from a template double-stranded polynucleotide comprising the nucleotide sequence of this invention, wherein the template double-stranded polynucleotide has been cleaved into double-stranded-random fragments of a desired size, and comprises the steps of adding to the resultant population of double-stranded random fragments one or more single or double-stranded oligonucleotides, wherein said oligonucleotides comprise an area of identity and an area of heterology to the double-stranded template polynucleotide; denaturing the resultant mixture of double-stranded random fragments and oligonucleotides into single-stranded fragments; incubating the resultant population of single-stranded fragments with a polymerase under conditions which result in the annealing of said single-stranded

fragments at said areas of identity to form pairs of annealed fragments, said areas of identity being sufficient for one member of a pair to prime replication of the other, thereby forming a mutagenized double-stranded polynucleotide; and repeating the second and third steps for at least two further cycles, wherein the resultant mixture in the second step of a further cycle includes the mutagenized double-stranded polynucleotide from the third step of the previous cycle, and the further cycle forms a further mutagenized double-stranded polynucleotide. In a preferred embodiment, the concentration of a single species of double-stranded random fragment in the population of double-stranded random fragments is less than 1% by weight of the total DNA. In a further preferred embodiment, the template double-stranded polynucleotide comprises at least about 100 species of polynucleotides. In another embodiment, the size of the double-stranded random fragments is from about 5 bp to 5 kb. In a further embodiment, the fourth step of the method comprises repeating the second and the third steps for at least 10 cycles.

The present invention also encompasses cells comprising a DNA molecule of the present invention, wherein the DNA molecule is not in its natural cellular environment. In a preferred embodiment, such cells are plant cells. In another preferred embodiment, a DNA molecule of the present invention is expressible in such cells and is comprised in an expression cassette which allow their expression in such cells. In a preferred embodiment, the expression cassette is stably integrated into the DNA of such host cell. In another preferred embodiment, the expression cassette is comprised in a vector, which is capable of replication in the cell and remains in the cell as an extrachromosomal molecule.

The present invention also encompasses a plant comprising the plant cells described above. In a further embodiment, the DNA molecules of the present invention are expressible in the plant, and expression of any one of the DNA molecules of the present invention or of a portion thereof in transgenic plants confers resistance upon the transgenic plant against fungal pathogens. In a preferred embodiment, the fungal pathogens desirably infect living epidermal cells, more desirably the fungal pathogens are from the order Erysiphales, also known as powdery mildews, preferably from the genus *Erysiphe* and more preferably the fungal pathogen is *Erysiphe graminis*. The present invention therefore also encompasses transgenic plants made resistant to fungal pathogens by the expression of any one of the DNA molecules of the present invention or of a portion thereof.

Plants transformed in accordance with the present invention may be monocots or dicots and include, but are not limited to, maize, wheat, barley, rye, sweet potato, bean, pea, chicory, lettuce, cabbage, cauliflower, broccoli, turnip, radish, spinach, asparagus, onion, garlic, pepper, celery, squash, pumpkin, hemp, zucchini, apple, pear, quince, melon, plum, cherry, peach, nectarine, apricot, strawberry, grape, raspberry, blackberry, pineapple, avocado, papaya, mango, banana, soybean, tomato, sorghum, sugarcane, sugarbeet, sunflower, rapeseed, clover, tobacco, carrot, cotton, alfalfa, rice, potato, eggplant, cucumber, *Arabidopsis thaliana*, and woody plants such as coniferous and deciduous trees, especially maize, wheat, or sugarbeet.

Once a desired nucleotide sequence has been transformed into a particular plant species, it may be propagated in that species or moved into other varieties of the same species, particularly including commercial varieties, using traditional breeding techniques.

For their expression in transgenic plants, the DNA molecules may require modification and optimization. It is known in the art that all organisms have specific preferences for codon usage, and the codons in the nucleotide sequence comprised in the DNA molecules of the present invention can be changed to conform with specific plant preferences, while maintaining the amino acids encoded thereby. Furthermore, high expression in plants is best achieved from coding sequences which have at least 35% GC content, and preferably more than 45%. Nucleotide sequences which have low GC contents may express poorly due to the existence of ATTTA motifs which may destabilize messages, and AATAAA motifs which may cause inappropriate polyadenylation. Although preferred gene sequences may be adequately expressed in both monocotyledonous and dicotyledonous plant species, sequences can be modified to account for the specific codon preferences and GC content preferences of monocotyledons or dicotyledons as these preferences have been shown to differ (Murray *et al.* Nucl. Acids Res. 17: 477-498 (1989)). In addition, the nucleotide sequences are screened for the existence of illegitimate splice sites which cause message truncation. All changes required to be made within the nucleotide sequences such as those described above are made using well known techniques of site directed mutagenesis, PCR, and synthetic gene construction using the methods described in the published patent applications EP 0 385 962, EP 0 359 472 and WO 93/07278.

For efficient initiation of translation, sequences adjacent to the initiating methionine may require modification. For example, they can be modified by the inclusion of sequences known to be effective in plants. Joshi has suggested an appropriate consensus for plants (NAR 15: 6643-6653 (1987)) and Clontech suggests a further consensus translation initiator

(1993/1994 catalog, page 210). These consensus sequences are suitable for use with the nucleotide sequences of this invention. The sequences are incorporated into constructions comprising the nucleotide sequence, up to and including the ATG (whilst leaving the second amino acid unmodified), or alternatively up to and including the GTC subsequent to the ATG (with the possibility of modifying the second amino acid of the transgene).

The DNA molecules in transgenic plants are driven by a promoter shown to be functional in plants. The choice of promoter will vary depending on the temporal and spatial requirements for expression, and also depending on the target species. For the protection of plants against foliar pathogens, expression in leaves is preferred; for the protection of plants against ear pathogens, expression in inflorescences (*e.g.* spikes, panicles, cobs, etc.) is preferred; for protection of plants against root pathogens, expression in roots is preferred; for protection of seedlings against soil-borne pathogens, expression in roots and/or seedlings is preferred. In many cases, however, protection against more than one type of phytopathogen is sought, and thus expression in multiple tissues is desirable. Although many promoters from dicotyledons have been shown to be operational in monocotyledons and *vice versa*, ideally dicotyledonous promoters are selected for expression in dicotyledons, and monocotyledonous promoters for expression in monocotyledons. However, there is no restriction to the provenance of selected promoters; it is sufficient that they are operational in driving the expression of the DNA molecules in the desired cell.

Preferred promoters which are expressed constitutively include promoters derived from *Agrobacterium* opine synthase genes, *e.g.* the *nos* promoter, or a dual promoter from the *Agrobacterium* Ti plasmid (Velten et al. (1984) EMBO J. 3: 2723-2730), or viral promoters operable in plants, *e.g.* the CaMV 35S and 19S promoters, and promoters from genes encoding actin or ubiquitin. Another preferred promoter is a synthetic promoter, such as the Gelvin Super MAS promoter (Ni et al. (1995) Plant J. 7: 661-676). The DNA molecules of this invention can also be expressed under the regulation of promoters which are chemically regulated. This enables the protein conferring fungal diseases to be synthesized only when the crop plants are treated with the inducing chemicals. Preferred technology for chemical induction of gene expression is detailed in the published application EP 0 332 104 and US patent 5,614,395. A preferred promoter for chemical induction is the tobacco PR-1a promoter.

A preferred category of promoters is that which is wound inducible. Numerous promoters have been described which are expressed at wound sites and also at the sites of

phytopathogen infection. Ideally, such a promoter should only be active locally at the sites of infection, and in this way the protein controlling fungal diseases only accumulates in cells which need to synthesize it to kill the invading insect pest. Preferred promoters of this kind include those described by Stanford *et al.* Mol. Gen. Genet. 215: 200-208 (1989), Xu *et al.* Plant Molec. Biol. 22: 573-588 (1993), Logemann *et al.* Plant Cell 1: 151-158 (1989), Rohrmeier & Lehle, Plant Molec. Biol. 22: 783-792 (1993), Firek *et al.* Plant Molec. Biol. 22: 129-142 (1993), and Warner *et al.* Plant J. 3: 191-201 (1993).

Preferred tissue specific expression patterns include green tissue specific, root specific, stem specific, and flower specific. Promoters suitable for expression in green tissue include many which regulate genes involved in photosynthesis and many of these have been cloned from both monocotyledons and dicotyledons. A preferred promoter is the maize PEPC promoter from the phosphoenol carboxylase gene (Hudspeth & Grula, Plant Molec. Biol. 12: 579-589 (1989)). A preferred promoter for root specific expression is that described by de Framond (FEBS 290: 103-106 (1991); EP 0 452 269) and a further preferred root-specific promoter is that from the T-1 gene provided by this invention. A preferred stem specific promoter is that described in US patent 5,625,136 and which drives expression of the maize *trpA* gene.

Preferred embodiments of the invention are transgenic plants expressing a DNA molecule in a root-specific fashion. Further preferred embodiments are transgenic plants expressing the DNA molecule in a wound-inducible or pathogen infection-inducible manner.

In addition to the selection of a suitable promoter, constructions for expression of the protein in plants require an appropriate transcription terminator to be attached downstream of the heterologous nucleotide sequence. Several such terminators are available and known in the art (*e.g.* *tm1* from CaMV, E9 from *rbcS*). Any available terminator known to function in plants can be used in the context of this invention.

Numerous other sequences can be incorporated into expression cassettes for the DNA molecules of this invention. These include sequences which have been shown to enhance expression such as intron sequences (*e.g.* from *Adh1* and *bronze1*) and viral leader sequences (*e.g.* from TMV, MCMV and AMV).

It may be preferable to target expression of the DNA molecules to different cellular localizations in the plant. In some cases, localization in the cytosol may be desirable, whereas in other cases, localization in some subcellular organelle may be preferred. Subcellular localization of transgene encoded enzymes can be undertaken using

techniques well known in the art. Typically, the DNA encoding the target peptide from a known organelle-targeted gene product is manipulated and fused upstream of the nucleotide sequence. Many such target sequences are known for the chloroplast and their functioning in heterologous constructions has been shown.

Vectors suitable for plant transformation are described elsewhere in this specification. For *Agrobacterium*-mediated transformation, binary vectors or vectors carrying at least one T-DNA border sequence are suitable, whereas for direct gene transfer any vector is suitable and linear DNA containing only the construction of interest may be preferred. In the case of direct gene transfer, transformation with a single DNA species or co-transformation can be used (Schocher *et al.* *Biotechnology* 4: 1093-1096 (1986)). For both direct gene transfer and *Agrobacterium*-mediated transfer, transformation is usually (but not necessarily) undertaken with a selectable marker which may provide resistance to an antibiotic (kanamycin, hygromycin or methatrexate) or a herbicide (glufosinate, glyphosate or a protoporphyrinogen oxidase inhibitor), or a selectable marker which may confer a selective advantage to transformed cells, such as a phospho-mannose isomerase gene. The choice of selectable marker is not, however, critical to the invention.

In another preferred embodiment, the DNA molecules of this invention are directly transformed into the plastid genome. Plastid transformation technology is extensively described in U.S. Patent Nos. 5,451,513, 5,545,817, and 5,545,818, in PCT application no. WO 95/16783, and in McBride *et al.* (1994) *Proc. Natl. Acad. Sci. USA* 91, 7301-7305. The basic technique for chloroplast transformation involves introducing regions of cloned plastid DNA flanking a selectable marker together with the DNA molecule of interest into a suitable target tissue, e.g., using biolistics or protoplast transformation (e.g., calcium chloride or PEG mediated transformation). The 1 to 1.5 kb flanking regions, termed targeting sequences, facilitate homologous recombination with the plastid genome and thus allow the replacement or modification of specific regions of the plastome. Initially, point mutations in the chloroplast 16S rRNA and rps12 genes conferring resistance to spectinomycin and/or streptomycin are utilized as selectable markers for transformation (Svab, Z., Hajdukiewicz, P., and Maliga, P. (1990) *Proc. Natl. Acad. Sci. USA* 87, 8526-8530; Staub, J. M., and Maliga, P. (1992) *Plant Cell* 4, 39-45). This resulted in stable homoplasmic transformants at a frequency of approximately one per 100 bombardments of target leaves. The presence of cloning sites between these markers allowed creation of a plastid targeting vector for introduction of foreign genes (Staub, J.M., and Maliga, P. (1993) *EMBO J.* 12, 601-606,

hereby incorporated by reference). Substantial increases in transformation frequency are obtained by replacement of the recessive rRNA or r-protein antibiotic resistance genes with a dominant selectable marker, the bacterial *aadA* gene encoding the spectinomycin-detoxifying enzyme aminoglycoside-3'-adenyltransferase (Svab, Z., and Maliga, P. (1993) *Proc. Natl. Acad. Sci. USA* **90**, 913-917). Previously, this marker had been used successfully for high-frequency transformation of the plastid genome of the green alga *Chlamydomonas reinhardtii* (Goldschmidt-Clermont, M. (1991) *Nucl. Acids Res.* **19**, 4083-4089). Other selectable markers useful for plastid transformation are known in the art and encompassed within the scope of the invention. Typically, approximately 15-20 cell division cycles following transformation are required to reach a homoplasmidic state. Plastid expression, in which genes are inserted by homologous recombination into all of the several thousand copies of the circular plastid genome present in each plant cell, takes advantage of the enormous copy number advantage over nuclear-expressed genes to permit expression levels that can readily exceed 10% of the total soluble plant protein.

The present invention also encompasses agricultural products, which comprise transgenic plants made resistant to fungal pathogens by expression of any one of the DNA molecules of the present invention or made resistant to fungal pathogens by any one of the methods described *infra*. Since such plants are resistant to fungal pathogens, pathogen growth on them is suppressed. Such plants and the agricultural products derived thereof are therefore less likely to contain mycotoxins, which are naturally produced by many fungal pathogens and which can be very toxic to human and to animals. Therefore, such agricultural products have better phytosanitary properties. In a preferred embodiment, such agricultural products are used as feedstuff, as silage, or as food.

It is a further object of the present invention, to provide methods of making plants resistant to fungal pathogens. The Mlo proteins encoded by the DNA molecules of the present invention confer upon a plant resistance to a fungal pathogen and it is a preferred object of the present invention to alter the expression of such proteins in their natural host environment. It is a further preferred object of the present invention to alter the stability or the activity of such proteins in their natural environment. Such alterations of the expression, stability, or activity of the proteins encoded by the DNA molecules of the present invention in a plant result in increased resistance of the plant to fungal pathogens. In a preferred embodiment, a protein encoded by a DNA molecule of the present invention the protein is a

negative regulator of resistance of a plant to fungal pathogens, in that it represses genetic pathways in the plant which are responsible for resistance of the plant to fungal pathogens. Therefore, it is a preferred object of the present invention to reduce the expression of the Mlo proteins encoded by the DNA molecules of the present invention in their natural host environment, or to reduce the stability or the activity of such proteins in their natural host environment.

"Sense" Suppression

In a preferred embodiment, reduction of the expression of a protein encoded by a DNA molecule of the present invention is obtained by "sense" suppression (referenced in e.g. Jorgensen et al. (1996) Plant Mol. Biol. 31, 957-973). In this case, the entirety or a portion of a DNA molecule of the present invention is comprised in an expression cassette, which is introduced in a host cell, preferably a plant cell, in which the DNA molecule is expressible. The DNA molecule is inserted in the expression cassette in the "sense orientation", meaning that the 5' end of the DNA molecule is adjacent to the promoter in the expression cassette and that the coding strand of the DNA molecule can be transcribed. In a preferred embodiment, the DNA molecule is fully translatable and all the genetic information comprised in the DNA molecule or a portion thereof is translated into a protein. In another preferred embodiment, the DNA molecule is partially translatable and a short peptide is translated. In a preferred embodiment, this is achieved by inserting at least one premature stop codon in the DNA molecule, which bring translation to a halt. In another more preferred embodiment, the DNA molecule is transcribed but no translation product is being made. This is usually achieved by removing the start codon, e.g. the "ATG", of the protein encoded by the DNA molecule. In a further preferred embodiment, the expression cassette comprising the DNA molecule or a portion thereof is stably integrated in the genome of the host cell. In another preferred embodiment, the expression cassette comprising the DNA molecule or a portion thereof is comprised in an extrachromosomal replicating molecule. In transgenic plants containing one of the expression cassettes described immediately above, the expression of the gene corresponding to the DNA molecule comprised in the expression cassette is reduced or abolished, leading to reduced levels of the protein or to its absence in the transgenic plants. As a result thereof, the transgenic plants are resistant to fungal pathogens.

"Anti-sense" Suppression

In another preferred embodiment, the reduction of the expression of a protein encoded by a DNA molecule of the present invention is obtained by "anti-sense" suppression. The entirety or a portion of a DNA molecule of the present invention is comprised in an expression cassette, wherein the DNA molecule is introduced in a host cell, preferably a plant cell, in which the DNA molecule is expressible. The DNA molecule is inserted in the expression cassette in the "anti-sense orientation", meaning that the 3' end of the DNA molecule is adjacent to the promoter in the expression cassette and that the non-coding strand of the DNA molecule can be transcribed. In a preferred embodiment, the expression cassette comprising the DNA molecule or a portion thereof is stably integrated in the genome of the host cell. In another preferred embodiment, the expression cassette comprising the DNA molecule or a portion thereof is comprised in an extrachromosomal replicating molecule. Several publications describing this approach are cited for further illustration (Green, P. J. et al., *Ann. Rev. Biochem.* 55:569-597 (1986); van der Krol, A. R. et al, *Antisense Nuc. Acids & Proteins*, pp. 125-141 (1991); Abel, P. P. et al., *Proc. Natl. Acad. Sci. USA* 86:6949-6952 (1989); Ecker, J. R. et al., *Proc. Natl. Acad. Sci. USA* 83:5372-5376 (Aug. 1986)).

Homologous Recombination

In another preferred embodiment, at least one genomic copy corresponding to a DNA molecule of the present invention is modified in the genome of the plant by homologous recombination as further illustrated in Paszkowski et al., *EMBO Journal* 7:4021-26 (1988). This technique uses the property of homologous sequences to recognize each other and to exchange nucleotide sequences between each by a process known in the art as homologous recombination. Homologous recombination can occur between the chromosomal copy of a nucleotide sequence in a cell and an incoming copy of the nucleotide sequence introduced in the cell by transformation. Specific modifications are thus accurately introduced in the chromosomal copy of the nucleotide sequence. In one embodiment, the regulatory elements of the gene encoding a protein of the present invention are modified. The existing regulatory elements are replaced by different regulatory elements, thus reducing expression of the protein, or they are mutated or deleted, thus abolishing the expression of the protein. In another embodiment, the coding region of the protein is modified by deletion of a part of the coding sequence of the entire coding sequence, or by mutation. Expression of a mutated protein can also confer upon the plant increased resistance to fungal pathogens.

In another preferred embodiment, a mutation in the chromosomal copy of a DNA molecule is introduced by transforming a cell with a chimeric oligonucleotide composed of a contiguous stretch of RNA and DNA residues in a duplex conformation with double hairpin caps on the ends. An additional feature of the oligonucleotide is the presence of 2'-O-methylation at the RNA residues. The RNA/DNA sequence is designed to align with the sequence of a chromosomal copy of a DNA molecule of the present invention and to contain the desired nucleotide change. This technique is further illustrated in US patent 5,501,967.

Ribozymes

In a further embodiment, the RNA coding for a protein of the present invention is cleaved by a catalytic RNA, or ribozyme, specific for such RNA. The ribozyme is expressed in transgenic plants and results in reduced amounts of RNA coding for the protein of the present invention in plant cells, thus leading to reduced amounts of protein accumulated in the cells and increased resistance of the plant to fungal pathogens. This method is further illustrated in US patent 4,987,071.

Dominant-Negative Mutants

In another preferred embodiment, the activity of the proteins encoded by the nucleotide sequences of this invention is changed. This is achieved by expression of dominant negative mutants of the proteins in transgenic plants, leading to the loss of activity of the endogenous protein.

Aptamers

In a further embodiment, the activity of a protein encoded by a DNA molecule of the present invention is inhibited by expressing in transgenic plants nucleic acid ligands, so-called aptamers, which specifically bind to the protein. Aptamers are preferentially obtained by the SELEX (Systematic Evolution of Ligands by EXponential Enrichment) method. In the SELEX method, a candidate mixture of single stranded nucleic acids having regions of randomized sequence is contacted with the protein and those nucleic acids having an increased affinity to the target are partitioned from the remainder of the candidate mixture. The partitioned nucleic acids are amplified to yield a ligand enriched mixture. After several iterations a nucleic acid with optimal affinity to the protein is obtained and is used for expression in transgenic plants. This method is further illustrated in US patent 5,270,163.

Methods for Isolating Nucleotide Sequences Comprising the Conserved Sequences

The conserved sequences comprised in Mlo proteins encoded by the DNA molecules of the present invention are used for the isolation of other DNA molecules encoding such sequences. In a preferred embodiment, a mixture of degenerated oligonucleotides that contain at least one possible oligonucleotide encoding a sequences set forth in SEQ ID No:1, or in SEQ ID No:2 is produced. The mixture of oligonucleotides encoding the sequences set forth in SEQ ID No:1 and the mixture of oligonucleotides complementary to sequences encoding the sequences set forth in SEQ ID No:2 are used in a PCR amplification reaction with a template DNA of choice. Mixtures of degenerated oligonucleotides are well known in the art and the degree of degeneration is varied as needed. In a preferred embodiment, the template DNA is a sample of total DNA from a plant, wherein such DNA sample is obtained by methods well known in the art. The amplified fragments resulting from the PCR reactions described above are isolated by methods well known in the art and are used to isolate the corresponding full-length cDNAs by screening a cDNA library or by using a RACE protocol, both well known in the art. This method represents a novel and useful strategy to isolate new genes conferring upon plants resistance to fungal pathogens.

The invention will be further described by reference to the following detailed examples. These examples are provided for purposes of illustration only, and are not intended to be limiting unless otherwise specified.

EXAMPLES

Example 1: Cloning and Sequencing of Mlo Genes from Wheat

Mlo genes from wheat are cloned using a reverse transcription-PCR approach. RNA is prepared from leaves of wheat cultivar UC703 and used to program reverse transcription using a Stratagene RT-PCR kit. The resulting cDNA is employed in PCR reactions using the following primers:

MLO-26 5' TTC CAG CAC CGG CAC AAG AA 3' (SEQ ID No:25)

MLO-10 5' AAG AAC TGC CTG AAG AAG GC 3' (SEQ ID No:23)
MLO-7 5' CAG AAA CTT GTC TCA TCC CTG G 3' (SEQ ID No:22)
MLO-5 5' ACA GAG ACC ACC TCC TTG GAA 3' (SEQ ID No:21)
MLO-15 5' CAC CAC CTT CAT GAT GCT CA 3' (SEQ ID No:24)

PCR is performed using the primer pairs listed below, the reactions of which resulted in amplification of fragments of the indicated sizes:

MLO-26 and MLO-10	503 bp
MLO-26 and MLO-7	1481 bp
MLO-5 and MLO-15	650 bp

The fragments are cloned into either pCR2.1 or pCR2.1-TOPO (Invitrogen). Plasmid DNA is prepared from the transformants and subjected to DNA sequencing. Sequencing reveals the existence of three different cDNA sequences with very high similarity to each other. These wheat Mlo genes are called TrMlo1, TrMlo2 and TrMlo3.

An additional wheat Mlo clone is isolated by screening a wheat cDNA library constructed in Lambda-ZAPII vector. To screen this library, mass excision is performed to convert the library into a Bluescript-based collection of cDNA clones. These are grown separately in pools of 80,000 independent clones, and plasmid DNA prepared. PCR reactions are performed on the pooled DNAs using oligonucleotide primers MLO-5 and MLO-15 (see above). Three pools yield a band of the expected size of 650 base pairs. These pools are subsequently fractionated by serial culturing of bacterial clones at lower and lower density, followed at each step by plasmid DNA preparation and PCR on each subpool using primers MLO-5 and MLO-15. After several rounds of fractionation a single clone is isolated which contained an insert with an Mlo sequence. Sequencing the inserts of two of these clones reveals that they contain identical inserts. The sequence of the third clone reveals an insert identical to that of the other two clones, but with 40 additional bases at the 5' end.

Cloning of the remaining portions of the wheat Mlo is accomplished by random amplification of cDNA ends (RACE). RACE reactions are performed on wheat UC703 poly-A⁺ RNA using a Marathon cDNA amplification kit (Clontech). Poly-A⁺ RNA is prepared from total

wheat RNA using an oligo-dT cellulose column (Gibco BRL). Oligonucleotides used for the RACE reactions are:

MLO-GSP1 5' TGG ACC TCT TCA TGT TCG ATC CCA TCT G 3' (SEQ ID No:26)

MLO-GSP2 5' CCT GAC GCT GTT CCA GAA TGC GTT TCA 3' (SEQ ID No:27)

Amplification using primer MLO-GSP1 and the 5' adapter primer provided in the kit results in a DNA fragment of ~1300 nucleotides. Amplification using primer MLO-GSP2 and the 3' adapter results in a DNA fragment of ~600 nucleotides. The fragments are cloned into pCR2.1-TOPO and called TrMlo1-5, TrMlo2-5 and TrMlo3-5 and comprise the 5' end of the wheat Mlo genes TrMlo1, TrMlo2 and TrMlo3, respectively. Plasmid DNA is prepared from the clones and the plasmid inserts are sequenced.

Example 2: Cloning of Mlo cDNAs from Arabidopsis

Comparison of the Mlo protein sequence to translations of database entries using the program TBLASTN revealed a number of entries with similarity to Mlo. Of these, full-length cDNAs corresponding to three Arabidopsis EST entries are cloned, with accession numbers H76041, N37544, and T22146. For each EST, oligonucleotides are designed to amplify a sequence corresponding to the EST from an Arabidopsis cDNA library constructed in plasmid pFL61 (Minet et al. (1992) Gene Nov 16; 121(2): 393-396.). The oligonucleotides used are:

N37544-1 5' AAG ATC AAG ATG AGG ACG TGG AAG TCG TGG 3' (SEQ ID No:29)

N37544-2 5' AGG CTG AAC CAC TGG GGC GCC TCT CAC CAC 3' (SEQ ID No:30)

T22146-1 5' CAA GTA TAT GAT GCG CGC TCT AGA GGA TGA 3' (SEQ ID No:31)

T22146-2 5' AGG TTT CAC CAC TAA GTC TCC TTC AAT GGC 3' (SEQ ID No:32)

H76041-1 5' GAT CAT TCA AGA CTT AGG CTC ACT CAT GAG 3' (SEQ ID No:33)

H76041-2 5' AAC AGC AAG GAA GAT TAC AAA TGA TGC CCA 3' (SEQ ID No:34)

The primers N37544-1 and N37544-2 amplify a ~500 base pair fragment from DNA prepared from the cDNA library, and the primers T22146-1 and T22146-2 amplify a ~250 base pair fragment. Primers H76041-1 and H76041-2 amplify two fragments of ~350 and a ~300 base pairs. The ~300 base fragment is of the size predicted from the EST sequence, and is used subsequently to diagnose the presence of a cDNA corresponding to the

H76041 EST in the cDNA library. DNA from the library is transformed into *E. coli*, and clones organized into pools of ~20,000 clones each. DNA from individual pools are screened by PCR using the different primer pairs, and positive pools subsequently subfractionated into smaller and smaller clone numbers. Isolation of individual positive clones is accomplished either by carrying this process through to completion, or in some cases by colony hybridization using EST sequences as probes once pool sizes has reached 200 clones or fewer. For ESTs N37544 and T22146, a clone corresponding to the EST is successfully isolated, and the insert sequenced. The cDNA corresponding to EST N37544 is designated CIB10295 in plasmid pCIB10295 and the plasmid containing the cDNA corresponding to EST T22146 (CIB10296) is designated pCIB10296. For these two ESTs, corresponding genomic sequences are available in the form of portions of Arabidopsis BAC clone sequences recently deposited to GenBank. Notably, the protein sequence predicted to be translated from these genomic sequences, as determined by GenBank, does not correspond to the sequence determined by directly sequencing of the cDNAs. Therefore, the amino acid sequence of the genes corresponding to the ESTs N37544 and T22146 are not obvious from the GenBank entry and are only elucidated through cloning and sequencing of the cDNA clones. The clone isolated using primers for the H76041 EST is found to contain not the gene for H76041, but a novel Mlo gene family member as insert. The insert is completely sequenced, and this Mlo gene family member is designated CIB10259 in plasmid pCIB10259.

Example 3: Construction of Vectors for Expression of the Mlo Genes in Wheat

Two vectors are constructed for "antisense" expression of barley Mlo gene in wheat. PCR is performed using barley cDNA and the primer pair MLO-5 and MLO-7 (see (1) above), and the reaction results in amplification of a 1124 bp fragment which is cloned into pGEM-T (Promega). This fragment is excised from pGEM-T using the enzymes SacII and NotI. The 1124 base pair fragment is cloned into pBluescript-SK(+). The insert is now excised with BamHI and SacI restriction sites and cloned into BamHI-SacI-digested pCIB9806 (described in patent application 08/838,219) in an orientation in which the Mlo coding sequence runs opposite to the maize ubiquitin promoter. This plasmid is designated pCK01.

To construct a vector for "antisense" expression of the entire Mlo gene in wheat, PCR is performed using the primer pair MLO-1 (5' ATG TCG GAC AAA AAA GGG GT 3' (SEQ ID No:19)) and MLO-10 (see (1) above), and the reaction results in amplification of a 635 bp fragment which is cloned into pCR2.1 (Invitrogen). This fragment is excised from pCR2.1

as an EcoRI fragment, and inserted into pGEM-9Zf(-) (Promega). A 320 nucleotide fragment spanning the naturally-occurring SacI site in Mlo to primer site MLO-10 is excised with SacI and BstXI. pCK01 is digested with SacI and BstXI, and the 320 base fragment inserted. To complete construction of the Mlo gene in the monocot expression vector, a 210 nucleotide SacI fragment is excised from the pGEM-9Zf(-) derivative. This fragment contains the 5' end of the Mlo coding sequence, from the primer site MLO-1 to the naturally-occurring SacI site in the Mlo gene. The pCK01 derivative is digested with SacI and the 210 base fragment inserted. Clones are analyzed for orientation of the 210 base fragment in the newly-constructed vector by PCR, using primers MLO-1 and MLO-10. Only clones in which the 210 base fragment is inserted in the antisense orientation relative to the ubiquitin promoter yielded a 530 base pair product corresponding to the 5' end of the Mlo coding sequence. The resulting plasmid contained the entire Mlo coding sequence in "antisense" orientation relative to the ubiquitin promoter, and is designated pCK02.

To construct a vector for expression of the Mlo gene in "sense" orientation, plasmid pCK02 is digested with BamHI to release the Mlo coding sequence as insert. The BamHI fragment is religated back into the pCK02 base vector. Colonies with the Mlo coding sequence in reverse orientation relative to pCK02 are identified by SacI digestion, which yields a 1.8 kb fragment in such clones, as opposed to a 210 base fragment in clones of identical configuration as pCK02. A clone with the Mlo coding sequence in "sense" orientation relative to the maize ubiquitin promoter is selected as designated pCK03.

Example 4: Construction of Vectors for Expression of the Mlo Genes in Arabidopsis

Mlo clones in pCIB10259, pCIB10295 and pCIB10296, along with pCK02 (for the barley Mlo gene) are used in PCR reactions, resulting in bands that carry the full-length gene sequences flanked by BamHI restriction sites. The sequences of the primers used are:

SAS-1: 5' GGA TTA AGA TCT AAT GGC 3' (SEQ ID No:35, for pCIB10295)

SAS-2: 5' CAA AGA TCT TCA TTT CTT AAA AG 3' (SEQ ID No:36, for pCIB10295)

SAS-3: 5' GCG GAT CCA TGT CGG ACA AAA AAG G 3' (SEQ ID No:37, for barley Mlo)

SAS-4: 5' GCG GAT CCT CAT CCC TGG CTG AAG G 3' (SEQ ID No:38, for barley Mlo)

SAS-5: 5' GGA TCC ACC ATG GCC ACA AGA TG 3' (SEQ ID No:39, for pCIB10259)

SAS-6: 5' GGA TCC TTA GTC AAT ATC ATT AGC 3' (SEQ ID No:40, for pCIB10259)

SAS-7: 5' GCG GAT CCA TGG GTC ACG GAG GAG AAG 3' (SEQ ID No:41, for pCIB10269)

SAS-8: 5' GCG GAT CCT CAG TTG TTA TGA TCA GGA 3' (SEQ ID No:42, for pCIB10296)

The bands are cloned into pCR2.1-TOPO, and the inserts sequenced from the resulting plasmids to confirm the absence of mutations introduced by PCR. The plasmids are digested with BamHI, and the inserts purified and cloned into BamHI-digested pPEH28, a shuttle vector containing a copy of the Arabidopsis ubiquitin gene promoter UBQ3 (Norris et al. (1993) Plant Molecular Biology 21: 895-906) immediately downstream of the BamHI site. Clones containing Mlo sequences fused to UBQ3 are identified, and restriction analysis performed to identify clones with the inserts in the "sense" and "antisense" orientations relative to UBQ3. For each Mlo gene, a clone with the insert in "sense" orientation and a clone with the insert in "antisense" orientation is digested with XbaI, and the insert purified and cloned into XbaI-digested pCIB200. This places the UBQ3-Mlo gene fusion between T-DNA borders.

Example 5: Transformation of Wheat and Identification of Expressors

Wheat is transformed by particle bombardment of immature embryos as described in detail in patent application WO 94/13822. Plantlets are regenerated on media containing Basta and subjected to PCR analysis. For diagnosis of the presence of the Mlo transgenes by PCR, the following primers are used:

MLO-3: 5' ATG CTA CCA CAC GCA GAT CG 3'
ST27: 5' ACT TCT GCA GGT CGA CTC TA 3'

The primer MLO-3 corresponds to a region of the Mlo transgene, while primer ST27 lies within the maize ubiquitin promoter sequence. The use of both Mlo gene and ubiquitin promoter primers in PCR eliminates false positives arising from the use of two Mlo primers, which might prime from the chromosomal copy of the Mlo gene present in wheat.

Plants confirmed to contain Mlo transgenes are subjected to RNA gel blot analysis, to determine whether they contain altered levels of chromosomally-encoded wheat Mlo mRNAs. Poly-A⁺ RNAs are prepared from individual transgenic lines, and blotted onto Hybond-N+ filters. Blots are probed with a 530 base fragment corresponding to the 5' end of the Mlo gene. This region is absent from the pCK01 clone; therefore, hybridization to antisense RNA expressed from the transgene in transgenic lines containing pCK01 does not occur. For pCK02 transgenic lines, in which the transgene contains this 5' end

fragment, the probe hybridizes to two bands of differing size. A ~2.5 kb mRNA corresponding to the antisense transgene is distinguished from the 2.0 kb mRNA derived from wheat chromosomally-encoded Mlo genes. The abundance of the 2.0 kb mRNA is monitored as a measurement of the efficiency of gene suppression achieved by the transgene in individual lines.

Example 6: Disease Testing of Transgenic Wheat Lines

Plants of transgenic and untransformed UC703 (control) wheat lines are grown in the greenhouse until they are two weeks old. Plants are moved to a Percival growth chamber (cycle of 8 hours dark, 16° C, and 16 hours light, 20° C), and inoculated with *Erysiphe graminis f. sp. tritici* by liberal application of spores. The degree of fungal sporulation is scored two weeks after inoculation. Plants are scored 1 (little to no hyphal growth and no visible sporulation), 2 (some hyphal growth and sporulation, but less control plants) or 3 (hyphal growth and sporulation comparable to controls). Transgenic wheat lines expressing Mlo-constructs show increased resistance to the pathogen. An example for results obtained with the antisense barley Mlo construct is shown below.

Screening of siblings from transgenic lines R1 and R2 for disease resistance

Siblings of Mlo antisense transformants (T2 seeds) are planted, inoculated with *E. graminis*, and scored for disease resistance.

	R1	R2	UC703 (control)
<u>Seed Planted</u>	24	24	24
Germinated	14	24	21
Disease Score 1	4	2	0
Disease Score 2	0	0	0
Disease Score 3	10	22	21

The fact that a small percentage of the R1 and R2 plants exhibit resistance may be due to the fact that T2 populations were tested which are still segregating for the transgene.

Example 7: Analysis of Arabidopsis Lines Expressing the Mlo Genes

Derivatives of pCIB200 containing Mlo genes are used to transform *Arabidopsis* ecotype Ws-O by vacuum infiltration (Bechtold, N., Ellis, J. and Pelletier, G. (1993) *C. R. Acad. Sci. Paris* 316, 1194-1199). Progeny are screened by kanamycin selection to identify transformants. For Mlo transgenic lines, plants are identified which expressed Mlo by RNA gel blot analysis. For Mlo genes, transformants are analyzed for alteration in the steady-state level of mRNA accumulation by RNA gel blot analysis. Transformants exhibiting sense or antisense suppression of target genes are assayed for alteration in reaction to the phytopathogenic fungi *Erysiphe cichoracearum* and *Peronospora parasitica*, and the bacterial pathogen *Pseudomonas syringae* pv. *tomato*. Leaves of transgenic plants are inspected both macroscopically and microscopically using trypan blue staining, to test for the presence of necrosis.

For *Erysiphe* inoculation, spores are liberally applied to *Arabidopsis* rosettes and plants are maintained in a Percival growth chamber at 25°C. Degree of fungal sporulation is scored 10 days after inoculation.

Example 8: Use of Regions of Similarity between Mlo Sequences to Isolate Additional Mlo Gene Family Members

Alignment of the amino acid sequences predicted to be encoded by Mlo genes revealed a number of short regions of high amino similarity between all the gene products. Degenerate primers are designed to these regions, and PCR reactions with these primers are carried out according to PCR reagent supplier's recommendations. The amplified fragments are used as a probe to isolate full-length cDNA or genomic clones of the novel Mlo genes. Amino acid sequences conserved between the Mlo proteins of the present invention (in bold) and degenerated oligonucleotides used for the isolation of additional Mlo genes are shown below:

	E	L	M	X1	X2	G	X3	I	S	L	L	L	X4
WHEAT													
TrMlo1	GAG	CTC	ATG	CTG	GTG	GGC	TTC	ATC					
TrMlo2	GAG	CTG	ATG	CTG	GTG	GGG	TTC	ATC					
TrMlo3	GAG	CTG	ATG	CTG	GTG	GGA	TTC	ATC					
ARABIDOPSIS													
CIB10259	GAG	CTG	ATG	ATT	CTA	GGA	TTC	ATT					

CIB10295 GAG CTT ATG CTG TTG GGA TTC ATA
 CIB10296 GAG CTG ATG TTG TTA GGG TTT ATA
 F19850 GAG CTG ATG GTT CTT GGA TTC ATC
 U95973 GAG TTG ATG TTG CTG GGA CTT ATA

5' GAG CTB ATG MTB BTR GGM TTC AT 3'

	X5	T	X6	P	L	X7	X8	X9	V	X10	Q	M	G	S
WHEAT														
TrM1o1							GCG	CTC	GTC	ACA	CAG	ATG	GGA	TCA
TrM1o2							GCG	CTC	GTC	ACA	CAG	ATG	GGA	TCG
TrM1o3							GCG	CTA	GTC	ACA	CAG	ATG	GGA	TCA

ARABIDOPSIS

CIB10259	GCA	CTA	GTT	ACT	CAG	ATG	GGT	TCA
CIB10295	GCA	CTT	GTT	ACT	CAG	ATG	GGT	AGT
CIB10296	GCC	ATC	GTC	TCA	CAG	ATG	GGA	AGT
F19850	GCA	CTC	GTA	ACT	CAG	ATG	GGT	TCT
U95973	GTA	ATC	GTT	ACT	CAG	ATG	GGA	TCT

5' WCC CAT CTG AGT GAC DAG BGC RTA 3'

X1= L, V or I, X2= V or L, X3= F or L, X4= T, S or A.

X5= I, V, S or G, X6= F, L or V, X7= Y or N, X8= A or V, X9= L or I, X10= T or S.

R=A,G Y=C,T M=A,C K=G,T S=C,G W=A,T H=A,C,T B=C,G,T V=A,C,G D=A,G,T

N=A,C,G,T.

Example 9: Modification of Coding Sequences and Adjacent Sequences

The DNA molecules described in this application can be modified for expression in transgenic plant hosts to achieve and optimize or down-regulate their expression. The following problems may be encountered and the modification of these DNA molecules can be undertaken using techniques well known in the art.

- (1) Codon Usage. The preferred codon usage in some plants differs from the preferred codon usage in some other plant species. Typically plant evolution has tended towards a strong preference of the nucleotides C and G in the third base position of monocotyledons.

whereas dicotyledons often use the nucleotides A or T at this position. By modifying a gene to incorporate preferred codon usage for a particular target transgenic species, many of the problems described below for GC/AT content and illegitimate splicing will be overcome.

(2) GC/AT Content. Plant genes typically have a GC content of more than 35%. DNA molecules which are rich in A and T nucleotides can cause several problems in plants. Firstly, motifs of ATTTA are believed to cause destabilization of messages and are found at the 3' end of many short-lived mRNAs. Secondly, the occurrence of polyadenylation signals such as AATAAA at inappropriate positions within the message is believed to cause premature truncation of transcription. In addition, monocotyledons may recognize AT-rich sequences as splice sites (see below).

(3) Sequences Adjacent to the Initiating Methionine. It is believed that ribosomes attach to the 5' end of the message and scan for the first available ATG at which to start translation. Nevertheless, it is believed that there is a preference for certain nucleotides adjacent to the ATG and that expression of DNA molecules of the present invention can be enhanced by the inclusion of a new consensus translation initiator at the ATG. Clontech (1993/1994 catalog, page 210) have suggested a sequence as a consensus translation initiator for the expression of the *E. coli uidA* gene in plants. Further, Joshi (NAR 15: 6643-6653 (1987)) has compared many plant sequences adjacent to the ATG and suggests a consensus sequence. In situations where difficulties are encountered in the expression of DNA molecules in plants, inclusion of one of these sequences at the initiating ATG may improve translation. In such cases the last three nucleotides of the consensus may not be appropriate for inclusion in the modified sequence due to their modification of the second AA residue. Preferred sequences adjacent to the initiating methionine may differ between different plant species. A survey of 14 maize genes located in the GenBank database provided the following results:

Position Before the Initiating ATG in 14 Maize Genes:

	<u>-10</u>	<u>-9</u>	<u>-8</u>	<u>-7</u>	<u>-6</u>	<u>-5</u>	<u>-4</u>	<u>-3</u>	<u>-2</u>	<u>-1</u>
C	3	8	4	6	2	5	6	0	10	7
T	3	0	3	4	3	2	1	1	1	0
A	2	3	1	4	3	2	3	7	2	3

G 6 3 6 0 6 5 4 6 1 5

This analysis can be done for the desired plant species into which the nucleotide sequence is being incorporated, and the sequence adjacent to the ATG modified to incorporate the preferred nucleotides.

(4) Removal of Illegitimate Splice Sites. DNA molecules of the present invention may also contain motifs which may be recognized in plants as 5' or 3' splice sites, and be cleaved, thus generating truncated or deleted messages. These sites can be removed using the techniques well known in the art.

(5) Creation of Dominant-Negative Mutants

In addition, DNA molecules of the present invention may also include molecules that are modified in such a way that the activity of the proteins encoded by the nucleotide sequences of this invention is changed. This is achieved by expression of dominant negative mutants of the proteins in transgenic plants, leading to the loss of activity of the endogenous protein. The location of mutations in the Mlo nucleotide sequence leading to the production of such dominant-negative mutations are listed below. Either a single mutation or a combination of the different mutations listed below can be introduced.

Mutation Number	Description of mutation	Location of mutation in wheat Mlo proteins of SEQ ID Nos: 4, 6 and 8 (aa)	Location of mutation in wheat Mlo genes (nt number)		
			TrMlo1 SEQ ID No:3	TrMlo2 SEQ ID No:5	TrMlo3 SEQ ID No:7
1	T to A	Trp (163) to Arg	662	487	684
2	deletion	frameshift after Pro (396)	1366-1367	1191-1192	1388-1389
3	deletion	frameshift after Trp (160)	656-666	481-491	678-688
4	G to A	Met (1) to Ile	178	3	200
5	G to A	Gly (227) to Asp	855	680	877

6	A to G	Met (1) to Val	176	1	198
7	A to T	Arg (11) to Trp	206	31	228
8	deletion	missing Phe (183), Thr (184)	721-726	546-551	743-748
9	T to A	Val (31) to Glu	267	92	289
10	C to T	Ser (32) to Phe	270	95	292
11	T to A	Leu (271) to His	987	812	1009

Techniques for the modification of coding sequences and adjacent sequences are well known in the art. In cases where the initial expression of a DNA molecule of the present invention is low and it is deemed appropriate to make alterations to the sequence as described above, then the construction of synthetic genes can be accomplished according to methods well known in the art. These are, for example, described in the published patent disclosures EP 0 385 962, EP 0 359 472 and WO 93/07278. In most cases it is preferable to assay the expression of gene constructions using transient assay protocols (which are well known in the art) prior to their transfer to transgenic plants.

Example 10: Construction of Plant Transformation Vectors

Numerous transformation vectors are available for plant transformation, and the DNA molecules of this invention can be used in conjunction with any such vectors. The selection of vector for use will depend upon the preferred transformation technique and the target species for transformation. For certain target species, different antibiotic or herbicide selection markers may be preferred. Selection markers used routinely in transformation include the *npfII* gene which confers resistance to kanamycin, paromomycin, geneticin and related antibiotics (Vieira and Messing, 1982, Gene 19: 259-268; Bevan et al., 1983, Nature 304:184-187) the bacterial *aadA* gene (Goldschmidt-Clermont, 1991, Nucl. Acids Res. 19: 4083-4089), encoding aminoglycoside 3'-adenylyltransferase and conferring resistance to streptomycin or spectinomycin, the *hph* gene which confers resistance to the antibiotic hygromycin (Blochliger and Diggelmann, 1984, Mol. Cell. Biol. 4: 2929-2931), and the *dhfr* gene, which confers resistance to methotrexate (Bourouis and Jarry, 1983, EMBO J. 2: 1099-1104). Other markers to be used include a phosphinothricin acetyltransferase gene, which confers resistance to the herbicide phosphinothricin (White et al., 1990, Nucl. Acids Res. 18: 1062; Spencer et al. 1990, Theor. Appl. Genet. 79: 625-631), a mutant EPSP

synthase gene encoding glyphosate resistance (Hinchee et al., 1988, Bio/Technology 6: 915-922), a mutant acetolactate synthase (ALS) gene which confers imidazolinone or sulfonylurea resistance (Lee et al., 1988, EMBO J. 7: 1241-1248), a mutant psbA gene conferring resistance to atrazine (Smeda et al., 1993, Plant Physiol. 103: 911-917), or a mutant protoporphyrinogen oxidase gene as described in US Patent No. 5,767, 373.

Selection markers resulting in positive selection, such as a phosphomannose isomerase gene, as described in US Patent No 5,767,378, are also used.

Identification of transformed cells may also be accomplished through expression of screenable marker genes such as genes coding for chloramphenicol acetyl transferase (CAT), β -glucuronidase (GUS), luciferase, and green fluorescent protein (GFP) or any other protein that confers a phenotypically distinct trait to the transformed cell.

(1) Construction of Vectors Suitable for *Agrobacterium* Transformation

Many vectors are available for transformation using *Agrobacterium tumefaciens*. These typically carry at least one T-DNA border sequence and include vectors such as pBIN19 (Bevan, Nucl. Acids Res. (1984)) and pXYZ. Below the construction of two typical vectors is described.

Construction of pCIB200 and pCIB2001

The binary vectors pCIB200 and pCIB2001 are used for the construction of recombinant vectors for use with *Agrobacterium* and is constructed in the following manner. pTJS75kan is created by *NarI* digestion of pTJS75 (Schmidhauser & Helinski, J Bacteriol. 164: 446-455 (1985)) allowing excision of the tetracycline-resistance gene, followed by insertion of an *AccI* fragment from pUC4K carrying an NPTII (Vieira & Messing, Gene 19: 259-268 (1982); Bevan et al., Nature 304: 184-187 (1983); McBride et al., Plant Molecular Biology 14: 266-276 (1990)). *XhoI* linkers are ligated to the *EcoRV* fragment of pCIB7 which contains the left and right T-DNA borders, a plant selectable *nos/nptII* chimeric gene and the pUC polylinker (Rothstein et al., Gene 53: 153-161 (1987)), and the *XhoI*-digested fragment is cloned into *Sall*-digested pTJS75kan to create pCIB200 (see also EP 0 332 104, example 19). pCIB200 contains the following unique polylinker restriction sites: *EcoRI*, *SstI*, *KpnI*, *BglII*, *XbaI*, and *Sall*. pCIB2001 is a derivative of pCIB200 which created by the insertion into the polylinker of additional restriction sites. Unique restriction sites in the polylinker of pCIB2001 are *EcoRI*, *SstI*, *KpnI*, *BglII*, *XbaI*, *Sall*, *MluI*, *BclI*, *AvrII*, *Apal*, *HpaI*, and *StuI*. pCIB2001, in addition to containing these unique restriction sites also has plant and

bacterial kanamycin selection, left and right T-DNA borders for *Agrobacterium*-mediated transformation, the RK2-derived *trfA* function for mobilization between *E. coli* and other hosts, and the *OriT* and *OriV* functions also from RK2. The pCIB2001 polylinker is suitable for the cloning of plant expression cassettes containing their own regulatory signals.

Construction of pCIB10 and Hygromycin Selection Derivatives thereof

The binary vector pCIB10 contains a gene encoding kanamycin resistance for selection in plants, T-DNA right and left border sequences and incorporates sequences from the wide host-range plasmid pRK252 allowing it to replicate in both *E. coli* and *Agrobacterium*. Its construction is described by Rothstein *et al.* (Gene 53: 153-161 (1987)). Various derivatives of pCIB10 have been constructed which incorporate the gene for hygromycin B phosphotransferase described by Gritz *et al.* (Gene 25: 179-188 (1983)). These derivatives enable selection of transgenic plant cells on hygromycin only (pCIB743), or hygromycin and kanamycin (pCIB715, pCIB717).

(2) Construction of Vectors Suitable for non-*Agrobacterium* Transformation.

Transformation without the use of *Agrobacterium tumefaciens* circumvents the requirement for T-DNA sequences in the chosen transformation vector and consequently vectors lacking these sequences can be utilized in addition to vectors such as the ones described above which contain T-DNA sequences. Transformation techniques which do not rely on *Agrobacterium* include transformation via particle bombardment, protoplast uptake (*e.g.* PEG and electroporation) and microinjection. The choice of vector depends largely on the preferred selection for the species being transformed. Below, the construction of some typical vectors is described.

Construction of pCIB3064

pCIB3064 is a pUC-derived vector suitable for direct gene transfer techniques in combination with selection by the herbicide Basta (or phosphinothricin). The plasmid pCIB246 comprises the CaMV 35S promoter in operational fusion to the *E. coli* GUS gene and the CaMV 35S transcriptional terminator and is described in the PCT published application WO 93/07278. The 35S promoter of this vector contains two ATG sequences 5' of the start site. These sites are mutated using standard PCR techniques in such a way as to remove the ATGs and generate the restriction sites *SspI* and *PvuII*. The new restriction sites are 96 and 37 bp away from the unique *Sall* site and 101 and 42 bp away from the actual start site. The resultant derivative of pCIB246 is designated pCIB3025. The GUS

gene is then excised from pCIB3025 by digestion with *Sall* and *SacI*, the termini rendered blunt and religated to generate plasmid pCIB3060. The plasmid pJIT82 is obtained from the John Innes Centre, Norwich and the a 400 bp *SmaI* fragment containing the *bar* gene from *Streptomyces viridochromogenes* is excised and inserted into the *HpaI* site of pCIB3060 (Thompson *et al.* EMBO J 6: 2519-2523 (1987)). This generated pCIB3064 which comprises the *bar* gene under the control of the CaMV 35S promoter and terminator for herbicide selection, a gene for ampicillin resistance (for selection in *E. coli*) and a polylinker with the unique sites *SphI*, *PstI*, *HindIII*, and *BamHI*. This vector is suitable for the cloning of plant expression cassettes containing their own regulatory signals.

Construction of pSOG19 and pSOG35

pSOG35 is a transformation vector which utilizes the *E. coli* gene dihydrofolate reductase (DHFR) as a selectable marker conferring resistance to methotrexate. PCR is used to amplify the 35S promoter (~800 bp), intron 6 from the maize *Adh1* gene (~550 bp) and 18 bp of the GUS untranslated leader sequence from pSOG10. A 250 bp fragment encoding the *E. coli* dihydrofolate reductase type II gene is also amplified by PCR and these two PCR fragments are assembled with a *SacI-PstI* fragment from pBI221 (Clontech) which comprised the pUC19 vector backbone and the nopaline synthase terminator. Assembly of these fragments generated pSOG19 which contains the 35S promoter in fusion with the intron 6 sequence, the GUS leader, the DHFR gene and the nopaline synthase terminator. Replacement of the GUS leader in pSOG19 with the leader sequence from Maize Chlorotic Mottle Virus (MCMV) generated the vector pSOG35. pSOG19 and pSOG35 carry the pUC gene for ampicillin resistance and have *HindIII*, *SphI*, *PstI* and *EcoRI* sites available for the cloning of foreign sequences.

Example 11: Requirements for Construction of Plant Expression Cassettes

Gene sequences intended for expression in transgenic plants are firstly assembled in expression cassettes behind a suitable promoter and upstream of a suitable transcription terminator.

Promoter Selection

The selection of promoter used in expression cassettes will determine the spatial and temporal expression pattern of the transgene in the transgenic plant. Selected promoters will express transgenes in specific cell types (such as leaf epidermal cells, mesophyll cells, root cortex cells) or in specific tissues or organs (roots, leaves or flowers, for example) and

this selection will reflect the desired location of biosynthesis of a DNA molecule of the present invention. Alternatively, the selected promoter may drive expression of the gene under a light-induced or other temporally regulated promoter. A further alternative is that the selected promoter be chemically regulated. This provides the possibility of inducing the expression of the nucleotide sequence only when desired and caused by treatment with a chemical inducer.

Transcriptional Terminators

A variety of transcriptional terminators are available for use in expression cassettes. These are responsible for the termination of transcription beyond the transgene and its correct polyadenylation. Appropriate transcriptional terminators and those which are known to function in plants and include the CaMV 35S terminator, the *tm1* terminator, the nopaline synthase terminator, the pea *rbcS* E9 terminator. These can be used in both monocotyledons and dicotyledons.

Sequences for the Enhancement or Regulation of Expression

Numerous sequences have been found to enhance gene expression from within the transcriptional unit and these sequences can be used in conjunction with the genes of this invention to increase their expression in transgenic plants.

Various intron sequences have been shown to enhance expression, particularly in monocotyledonous cells. For example, the introns of the maize *Adh1* gene have been found to significantly enhance the expression of the wild-type gene under its cognate promoter when introduced into maize cells. Intron 1 is found to be particularly effective and enhanced expression in fusion constructs with the chloramphenicol acetyltransferase gene (Callis *et al.*, Genes Develop **1**: 1183-1200 (1987)). In the same experimental system, the intron from the maize *bronze1* gene had a similar effect in enhancing expression (Callis *et al.*, *supra*). Intron sequences have been routinely incorporated into plant transformation vectors, typically within the non-translated leader.

A number of non-translated leader sequences derived from viruses are also known to enhance expression, and these are particularly effective in dicotyledonous cells. Specifically, leader sequences from Tobacco Mosaic Virus (TMV, the 'Ω-sequence'), Maize Chlorotic Mottle Virus (MCMV), and Alfalfa Mosaic Virus (AMV) have been shown to be effective in enhancing expression (*e.g.* Gallie *et al.* Nucl. Acids Res. **15**: 8693-8711 (1987); Skuzeski *et al.* Plant Molec. Biol. **15**: 65-79 (1990))

Targeting of the Gene Product Within the Cell

Various mechanisms for targeting gene products are known to exist in plants and the sequences controlling the functioning of these mechanisms have been characterized in some detail. For example, the targeting of gene products to the chloroplast is controlled by a signal sequence found at the aminoterminal end of various proteins and which is cleaved during chloroplast import yielding the mature protein (*e.g.* Comai *et al.* J. Biol. Chem. 263: 15104-15109 (1988)). These signal sequences can be fused to heterologous gene products to effect the import of heterologous products into the chloroplast (van den Broeck *et al.* Nature 313: 358-363 (1985)). DNA encoding for appropriate signal sequences can be isolated from the 5' end of the cDNAs encoding the RUBISCO protein, the CAB protein, the EPSP synthase enzyme, the GS2 protein and many other proteins which are known to be chloroplast localized.

Other gene products are localized to other organelles such as the mitochondrion and the peroxisome (*e.g.* Unger *et al.* Plant Molec. Biol. 13: 411-418 (1989)). The cDNAs encoding these products can also be manipulated to effect the targeting of heterologous gene products to these organelles. Examples of such sequences are the nuclear-encoded ATPases and specific aspartate amino transferase isoforms for mitochondria. Targeting to cellular protein bodies has been described by Rogers *et al.* (Proc. Natl. Acad. Sci. USA 82: 6512-6516 (1985)).

In addition sequences have been characterized which cause the targeting of gene products to other cell compartments. Aminoterminal sequences are responsible for targeting to the ER, the apoplast, and extracellular secretion from aleurone cells (Koehler & Ho, Plant Cell 2: 769-783 (1990)). Additionally, aminoterminal sequences in conjunction with carboxyterminal sequences are responsible for vacuolar targeting of gene products (Shinshi *et al.* Plant Molec. Biol. 14: 357-368 (1990)).

By the fusion of the appropriate targeting sequences described above to transgene sequences of interest it is possible to direct the transgene product to any organelle or cell compartment. For chloroplast targeting, for example, the chloroplast signal sequence from the RUBISCO gene, the CAB gene, the EPSP synthase gene, or the GS2 gene is fused in frame to the aminoterminal ATG of the transgene. The signal sequence selected should include the known cleavage site and the fusion constructed should take into account any amino acids after the cleavage site which are required for cleavage. In some cases this requirement may be fulfilled by the addition of a small number of amino acids between the cleavage site and the transgene ATG or alternatively replacement of some amino acids within the transgene sequence. Fusions constructed for chloroplast import can be tested

for efficacy of chloroplast uptake by *in vitro* translation of *in vitro* transcribed constructions followed by *in vitro* chloroplast uptake using techniques described by (Bartlett *et al.* In: Edelman *et al.* (Eds.) Methods in Chloroplast Molecular Biology, Elsevier. pp 1081-1091 (1982); Wasmann *et al.* Mol. Gen. Genet. 205: 446-453 (1986)). These construction techniques are well known in the art and are equally applicable to mitochondria and peroxisomes. The choice of targeting which may be required for the insecticidal toxin. This will usually be cytosolic or chloroplastic, although it may in some cases be mitochondrial or peroxisomal. The expression of the nucleotide sequence may also require targeting to the ER, the apoplast or the vacuole.

The above described mechanisms for cellular targeting can be utilized not only in conjunction with their cognate promoters, but also in conjunction with heterologous promoters so as to effect a specific cell targeting goal under the transcriptional regulation of a promoter which has an expression pattern different to that of the promoter from which the targeting signal derives.

Example 12: Examples of Expression Cassette Construction

The present invention encompasses the expression of a DNA molecule under the regulation of any promoter which is expressible in plants, regardless of the origin of the promoter. Furthermore, the invention encompasses the use of any plant-expressible promoter in conjunction with any further sequences required or selected for the expression of the DNA molecule. Such sequences include, but are not restricted to, transcriptional terminators, extraneous sequences to enhance expression (such as introns [*e.g.* *Adh* intron 1], viral sequences [*e.g.* TMV- Ω]), and sequences intended for the targeting of the gene product to specific organelles and cell compartments.

Constitutive Expression: the CaMV 35S Promoter

Construction of the plasmid pCGN1761 is described in the published patent application EP 0 392 225. pCGN1761 contains the 'double' 35S promoter and the *tm1* transcriptional terminator with a unique *EcoRI* site between the promoter and the terminator and has a pUC-type backbone. A derivative of pCGN1761 is constructed which has a modified polylinker which includes *NotI* and *XhoI* sites in addition to the existing *EcoRI* site. This derivative is designated pCGN1761ENX. pCGN1761ENX is useful for the cloning of cDNA sequences or gene sequences (including microbial ORF sequences) within its polylinker for the purposes of their expression under the control of the 35S promoter in transgenic plants.

The entire 35S promoter-gene sequence-*tml* terminator cassette of such a construction can be excised by *HindIII*, *SphI*, *Sall*, and *XbaI* sites 5' to the promoter and *XbaI*, *BamHI* and *BglII* sites 3' to the terminator for transfer to transformation vectors such as those described above in example 35. Furthermore, the double 35S promoter fragment can be removed by 5' excision with *HindIII*, *SphI*, *Sall*, *XbaI*, or *PstI*, and 3' excision with any of the polylinker restriction sites (*EcoRI*, *NotI* or *XhoI*) for replacement with another promoter.

Modification of pCGN1761ENX by Optimization of the Translational Initiation Site

For any of the constructions described in this section, modifications around the cloning sites can be made by the introduction of sequences which may enhance translation. This is particularly useful when genes derived from microorganisms are to be introduced into plant expression cassettes as these genes may not contain sequences adjacent to their initiating methionine which may be suitable for the initiation of translation in plants. In cases where genes derived from microorganisms are to be cloned into plant expression cassettes at their ATG it may be useful to modify the site of their insertion to optimize their expression. Modification of pCGN1761ENX is described by way of example to incorporate one of several optimized sequences for plant expression (*e.g.* Joshi, *supra*).

Expression under a Chemically Regulatable Promoter

This section describes the replacement of the double 35S promoter in pCGN1761ENX with any promoter of choice; by way of example the chemically regulated PR-1a promoter is described. The promoter of choice is preferably excised from its source by restriction enzymes, but can alternatively be PCR-amplified using primers which carry appropriate terminal restriction sites. Should PCR-amplification be undertaken, then the promoter should be resequenced to check for amplification errors after the cloning of the amplified promoter in the target vector. The chemically regulatable tobacco PR-1a promoter is cleaved from plasmid pCIB1004 (see EP 0 332 104, example 21 for construction) and transferred to plasmid pCGN1761ENX. pCIB1004 is cleaved with *NcoI* and the resultant 3' overhang of the linearized fragment is rendered blunt by treatment with T4 DNA polymerase. The fragment is then cleaved with *HindIII* and the resultant PR-1a promoter containing fragment is gel purified and cloned into pCGN1761ENX from which the double 35S promoter has been removed. This is done by cleavage with *XhoI* and blunting with T4 polymerase, followed by cleavage with *HindIII* and isolation of the larger vector-terminator containing fragment into which the pCIB1004 promoter fragment is cloned. This generates a pCGN1761ENX derivative with the PR-1a promoter and the *tml* terminator and an

intervening polylinker with unique *EcoRI* and *NotI* sites. The DNA molecule of the present invention can be inserted into this vector, and the fusion products (*i.e.* promoter-gene-terminator) can subsequently be transferred to any selected transformation vector, including those described in this application.

Constitutive Expression: the Actin Promoter

Several isoforms of actin are known to be expressed in most cell types and consequently the actin promoter is a good choice for a constitutive promoter. In particular, the promoter from the rice *Act1* gene has been cloned and characterized (McElroy *et al.* Plant Cell **2**: 163-171 (1990)). A 1.3 kb fragment of the promoter is found to contain all the regulatory elements required for expression in rice protoplasts. Furthermore, numerous expression vectors based on the *Act1* promoter have been constructed specifically for use in monocotyledons (McElroy *et al.* Mol. Gen. Genet. **231**: 150-160 (1991)). These incorporate the *Act1*-intron 1, *Adh1* 5' flanking sequence and *Adh1*-intron 1 (from the maize alcohol dehydrogenase gene) and sequence from the CaMV 35S promoter. Vectors showing highest expression are fusions of 35S and the *Act1* intron or the *Act1* 5' flanking sequence and the *Act1* intron. Optimization of sequences around the initiating ATG (of the GUS reporter gene) also enhanced expression. The promoter expression cassettes described by McElroy *et al.* (Mol. Gen. Genet. **231**: 150-160 (1991)) can be easily modified for the expression of the DNA molecules of the present invention and are particularly suitable for use in monocotyledonous hosts. For example, promoter containing fragments can be removed from the McElroy constructions and used to replace the double 35S promoter in pCGN1761ENX, which is then available for the insertion of specific gene sequences. The fusion genes thus constructed can then be transferred to appropriate transformation vectors. In a separate report the rice *Act1* promoter with its first intron has also been found to direct high expression in cultured barley cells (Chibbar *et al.* Plant Cell Rep. **12**: 506-509 (1993)).

Constitutive Expression: the Ubiquitin Promoter

Ubiquitin is another gene product known to accumulate in many cell types and its promoter has been cloned from several species for use in transgenic plants (*e.g.* sunflower - Binet *et al.* Plant Science **79**: 87-94 (1991), maize - Christensen *et al.* Plant Molec. Biol. **12**: 619-632 (1989)). The maize ubiquitin promoter has been developed in transgenic monocot systems and its sequence and vectors constructed for monocot transformation are disclosed in the patent publication EP 0 342 926. Further, Taylor *et al.* (Plant Cell Rep. **12**: 491-495 (1993))

describe a vector (pAHC25) which comprises the maize ubiquitin promoter and first intron and its high activity in cell suspensions of numerous monocotyledons when introduced via microprojectile bombardment. The ubiquitin promoter is clearly suitable for the expression of a DNA molecule of the present invention in transgenic plants, especially monocotyledons. Suitable vectors are derivatives of pAHC25 or any of the transformation vectors described in this application, modified by the introduction of the appropriate ubiquitin promoter and/or intron sequences.

Root Specific Expression

A preferred pattern of expression for the nucleotide sequence of the instant invention is root expression. Root expression is particularly useful for the control of soil-borne fungal pathogens. A suitable root promoter is that described by de Framond (FEBS 290: 103-106 (1991)) and also in the published patent application EP 0 452 269. This promoter is transferred to a suitable vector such as pCGN1761ENX for the insertion of the nucleotide sequence and subsequent transfer of the entire promoter-gene-terminator cassette to a transformation vector of interest.

Wound Inducible Promoters

Wound-inducible promoters are particularly suitable for the expression of a DNA molecules of the present invention because they are typically active not just on wound induction, but also at the sites of phytopathogen infection. Numerous such promoters have been described (*e.g.* Xu *et al.* Plant Molec. Biol. 22: 573-588 (1993), Logemann *et al.* Plant Cell 1: 151-158 (1989), Rohrmeier & Lehle, Plant Molec. Biol. 22: 783-792 (1993), Firek *et al.* Plant Molec. Biol. 22: 129-142 (1993), Warner *et al.* Plant J. 3: 191-201 (1993)) and all are suitable for use with the instant invention. Logemann *et al.* (*supra*) describe the 5' upstream sequences of the dicotyledonous potato *wun1* gene. Xu *et al.* (*supra*) show that a wound inducible promoter from the dicotyledon potato (*pin2*) is active in the monocotyledon rice. Further, Rohrmeier & Lehle (*supra*) describe the cloning of the maize *Wip1* cDNA which is wound induced and which can be used to isolated the cognate promoter using standard techniques. Similarly, Firek *et al.* (*supra*) and Warner *et al.* (*supra*) have described a wound induced gene from the monocotyledon *Asparagus officinalis* which is expressed at local wound and pathogen invasion sites. Using cloning techniques well known in the art, these promoters can be transferred to suitable vectors, fused to the DNA molecule of this invention, and used to express these genes at the sites of phytopathogen infection.

Pith Preferred Expression

Patent Application WO 93/07278 (to Ciba-Geigy) describes the isolation of the maize *tpa* gene which is preferentially expressed in pith cells. The gene sequence and promoter extend up to -1726 from the start of transcription are presented. Using standard molecular biological techniques, this promoter or parts thereof, can be transferred to a vector such as pCGN1761 where it can replace the 35S promoter and be used to drive the expression of a DNA molecule of this invention in a pith-preferred manner. In fact fragments containing the pith-preferred promoter or parts thereof can be transferred to any vector and modified for utility in transgenic plants.

Pollen-Specific Expression

Patent Application WO 93/07278 (to Ciba-Geigy) further describes the isolation of the maize calcium-dependent protein kinase (CDPK) gene which is expressed in pollen cells. The gene sequence and promoter extend up to 1400 bp from the start of transcription. Using standard molecular biological techniques, this promoter or parts thereof, can be transferred to a vector such as pCGN1761 where it can replace the 35S promoter and be used to drive the expression of a DNA molecule of this invention. In fact fragments containing the pollen-specific promoter or parts thereof can be transferred to any vector and modified for utility in transgenic plants.

Leaf-Specific Expression

A maize gene encoding phosphoenol carboxylase (PEPC) has been described by Hudspeth & Grula (Plant Molec Biol 12: 579-589 (1989)). Using standard molecular biological techniques the promoter for this gene can be used to drive the expression of any gene in a leaf-specific manner in transgenic plants.

Expression with Chloroplast Targeting

Chen & Jagendorf (J. Biol. Chem. 268: 2363-2367 (1993) have described the successful use of a chloroplast transit peptide for import of a heterologous transgene. This peptide used is the transit peptide from the *rbcS* gene from *Nicotiana plumbaginifolia* (Poulsen *et al.* Mol. Gen. Genet. 205: 193-200 (1986)). Using the restriction enzymes *DraI* and *SphI*, or *Tsp509I* and *SphI* the DNA sequence encoding this transit peptide can be excised from plasmid prbcS-8B (Poulsen *et al. supra*) and manipulated for use with any of the constructions described above. The *DraI-SphI* fragment extends from -58 relative to the initiating *rbcS* ATG to, and including, the first amino acid (also a methionine) of the mature peptide immediately after the import cleavage site, whereas the *Tsp509I-SphI* fragment

extends from -8 relative to the initiating *rbcS* ATG to, and including, the first amino acid of the mature peptide. Thus, these fragment can be appropriately inserted into the polylinker of any chosen expression cassette generating a transcriptional fusion to the untranslated leader of the chosen promoter (e.g. 35S, PR-1a, actin, ubiquitin *etc.*), whilst enabling the insertion of the DNA molecule of this invention in correct fusion downstream of the transit peptide. Constructions of this kind are routine in the art. For example, whereas the *DraI* end is already blunt, the 5' *Tsp509I* site may be rendered blunt by T4 polymerase treatment, or may alternatively be ligated to a linker or adaptor sequence to facilitate its fusion to the chosen promoter. The 3' *SphI* site may be maintained as such, or may alternatively be ligated to adaptor or linker sequences to facilitate its insertion into the chosen vector in such a way as to make available appropriate restriction sites for the subsequent insertion of the DNA molecule of this invention. Ideally the ATG of the *SphI* site is maintained and comprises the first ATG of the DNA molecule of this invention. Chen & Jagendorf (*supra*) provide consensus sequences for ideal cleavage for chloroplast import, and in each case a methionine is preferred at the first position of the mature protein. At subsequent positions there is more variation and the amino acid may not be so critical. In any case, fusion constructions can be assessed for efficiency of import *in vitro* using the methods described by Bartlett *et al.* (In: Edelman *et al.* (Eds.) *Methods in Chloroplast Molecular Biology*, Elsevier. pp 1081-1091 (1982)) and Wasmann *et al.* (*Mol. Gen. Genet.* 205: 446-453 (1986)). Typically the best approach may be to generate fusions using the DNA molecule of this invention with no modifications at the aminotermminus, and only to incorporate modifications when it is apparent that such fusions are not chloroplast imported at high efficiency, in which case modifications may be made in accordance with the established literature (Chen & Jagendorf, *supra*; Wasman *et al.*, *supra*; Ko & Ko, *J. Biol. Chem.* 267: 13910-13916 (1992)).

Similar manipulations can be undertaken to utilize other GS2 chloroplast transit peptide encoding sequences from other sources (monocotyledonous and dicotyledonous) and from other genes. In addition, similar procedures can be followed to achieve targeting to other subcellular compartments such as mitochondria.

What is claimed is:

1. A DNA molecule encoding an Mlo protein, wherein said Mlo protein comprises at least one of the sequences set forth in SEQ ID No:1 or SEQ ID No:2 and wherein said Mlo protein confers upon a plant resistance to a fungal pathogen.
2. The DNA molecule of claim 1, wherein said DNA molecule is identical or substantially similar to any one of the nucleotide sequences set forth in SEQ ID No:3, SEQ ID No:5 or SEQ ID No:7, or encodes an Mlo protein identical or substantially similar to an Mlo protein set forth in SEQ ID No:4, SEQ ID No:6 or SEQ ID No:8.
3. The DNA molecule of claim 1, wherein said DNA molecule is identical or substantially similar to any one of the nucleotide sequences set forth in SEQ ID No:9, SEQ ID No:11, SEQ ID No:13, SEQ ID No:15 or SEQ ID No:17, or encodes an Mlo protein identical or substantially similar to an Mlo protein encoded set forth in SEQ ID No:10, SEQ ID No:12, SEQ ID No:14, SEQ ID No:16 or SEQ ID No:18.
4. The DNA molecule of any one of claims 1 to 3, wherein said DNA molecule is not derived from barley.
5. The DNA molecule of any one of claims 1 to 4, wherein said DNA is modified such that the activity of the endogenous protein is lost.
6. The DNA molecule of claim 5, wherein the DNA modification results in one, all or a combination of the following changes in the amino acid sequence of the corresponding protein
 - Trp (163) to Arg
 - frameshift after Pro (396)
 - frameshift after Trp (160)
 - Met (1) to Ile
 - Gly (227) to Asp
 - Met (1) to Val
 - Arg (11) to Trp

- missing Phe (183), Thr (184)
- Val (31) to Glu
- Ser (32) to Phe
- Leu (271) to His

7. A DNA molecule antisense to a DNA molecule of any one of claims 1 to 6.

8. A protein comprising at least one of the sequences set forth in SEQ ID No:1 or SEQ ID No:2, wherein said protein is an Mlo protein and confers upon a plant resistance to a fungal pathogen.

9. The protein of claim 8, wherein said protein is encoded by a nucleotide sequence identical or substantially similar to any one of the sequences set forth in SEQ ID No:3, SEQ ID No:5 or SEQ ID No:7, or is identical or substantially similar to any one of the proteins set forth in SEQ ID No:4, SEQ ID No:6 or SEQ ID No:8.

10. The protein of claim 8, wherein said protein is encoded by a nucleotide sequence identical or substantially similar to any one of the sequences set forth in SEQ ID No:9, SEQ ID No:11, SEQ ID No:13, SEQ ID No:15 or SEQ ID No:17, or is identical or substantially similar to any one of the proteins set forth in SEQ ID No:10, SEQ ID No:12, SEQ ID No:14, SEQ ID No:16 or SEQ ID No:18.

11. The protein of any one of claims 8 to 10, wherein said protein is not derived from barley.

12. An expression cassette comprising a DNA molecule of any one of claims 1 to 7.

13. A vector comprising an expression cassette comprising a DNA molecule of claim 12.

14. A cell comprising an expression cassette or parts of it comprising a DNA molecule of any one of claims 1 to 7, wherein said DNA molecule in said expression cassette is expressible in said cell.

15. The cell of claim 14, wherein said DNA molecule is stably integrated in the genome of said cell.
16. The cell of any one of claims 14 or 15, wherein said cell is a plant cell.
17. A plant comprising an expression cassette or parts of it comprising a DNA molecule of any one of claims 1 to 7, wherein said DNA molecule in said expression cassette is expressible in said plant.
18. The plant of claim 17, wherein said DNA molecule is stably integrated in the plant genome of said plant.
19. An agricultural product comprising a plant comprising an isolated DNA molecule of any one of claims 1 to 7, wherein said agricultural product has improved phytosanitary properties.
20. A method for making a plant resistant to a fungal pathogen comprising the steps of:
- a) expressing in said plant a DNA molecule of any one of claims 1 to 6 in "sense" orientation; or
 - b) expressing in said plant a DNA molecule of any one of claims 1 to 6 in "anti-sense" orientation; or
 - c) expressing in said plant a ribozyme capable of specifically cleaving a messenger RNA transcript encoded by an endogenous gene corresponding to a DNA molecule of any one of claims 1 to 6; or
 - d) expressing in a plant an aptamer specifically directed to a protein or part of a protein encoded by a DNA molecule of any one of claims 1 to 6; or
 - e) expressing in a plant a mutated or a truncated form of a DNA molecule of any one of claims 1 to 6; or
 - f) modifying by homologous recombination in a plant at least one chromosomal copy of the gene corresponding to a DNA molecule of any one of claims 1 to 6.
21. A plant made resistant to a fungal pathogen by the method of claim 20.

22. The plant of claim 21, wherein said fungal pathogen infects living epidermal cells.
23. The plant of claim 21, wherein said fungal pathogen is from the order Erysiphales.
24. The plant of claim 21, wherein said fungal pathogen is from the genus *Erysiphe*.
25. The plant of claim 21, wherein said fungal pathogen is *Erysiphe graminis*.
26. An agricultural product with improved phytosanitary properties obtained using the method of claim 20.
27. A method for isolating DNA molecules encoding Mlo proteins comprising the steps of:
- a) mixing a degenerated oligonucleotide encoding at least six amino acids of SEQ ID No:1 and a degenerated oligonucleotide complementary to a sequence encoding at least six amino acids of SEQ ID No:2, with DNA extracted from a plant under conditions allowing hybridization of said degenerated oligonucleotides to said DNA; and
 - b) amplifying a DNA fragment of said plant DNA, wherein said DNA fragment comprises at its left and right ends nucleotide sequences that can anneal to said degenerated oligonucleotides in step a); and
 - c) obtaining a full-length cDNA clone comprising the DNA fragment of step b).
28. A method for mutagenizing a DNA molecule of claim 1, wherein said DNA molecule has been cleaved into double-stranded-random fragments of a desired size, and comprising the steps of:
- a) adding to the resultant population of double-stranded random fragments one or more single or double-stranded oligonucleotides, wherein said oligonucleotides comprise an area of identity and an area of heterology to the double-stranded template polynucleotide;
 - b) denaturing the resultant mixture of double-stranded random fragments and oligonucleotides into single-stranded fragments;
 - c) incubating the resultant population of single-stranded fragments with a polymerase under conditions which result in the annealing of said single-stranded fragments at said areas of identity to form pairs of annealed fragments, said areas of identity being

sufficient for one member of a pair to prime replication of the other, thereby forming a mutagenized double-stranded polynucleotide; and

- d) repeating the second and third steps for at least two further cycles, wherein the resultant mixture in the second step of a further cycle includes the mutagenized double-stranded polynucleotide from the third step of the previous cycle, and the further cycle forms a further mutagenized double-stranded polynucleotide.

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cagacgaccg gcggtctgggt ctccggacaa ggaaagaggt tgcgctcggg gaccg atg 178
                                         Met
                                         1

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Ala Asp Asp Asp Glu Tyr Pro Pro Ala Arg Thr Leu Pro Glu Thr Pro
          5                      10                      15

tcc tgg gcg gtg gcc ctc gtc ttc gcc gtc atg atc atc gtg tcc gtc 274
Ser Trp Ala Val Ala Leu Val Phe Ala Val Met Ile Ile Val Ser Val
          20                      25                      30

ctc ctg gag cac gcg ctc cat aag ctc ggc cat tgg ttc cac aag cgg 322
Leu Leu Glu His Ala Leu His Lys Leu Gly His Trp Phe His Lys Arg
          35                      40                      45

cac aag aac gcg ctg gcg gag gcg ctg gag aag atc aag gcg gag ctc 370
His Lys Asn Ala Leu Ala Glu Ala Leu Glu Lys Ile Lys Ala Glu Leu
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atg ctg gtg ggc ttc atc tcg ctg ctg ctc gcc gtg acg cag gac ccc 418
Met Leu Val Gly Phe Ile Ser Leu Leu Leu Ala Val Thr Gln Asp Pro
          70                      75                      80

atc tcc ggg ata tgc atc tcc gag aag gcc gcc agc atc atg cgg ccc 466
Ile Ser Gly Ile Cys Ile Ser Glu Lys Ala Ala Ser Ile Met Arg Pro
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aaa tgg gag aca gag acc gcc tcc ctg gaa tac cag ttc gca aat gat				706
Lys Trp Glu Thr Glu Thr Ala Ser Leu Glu Tyr Gln Phe Ala Asn Asp				
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Ala Gly Phe Ile Asn Ala His Leu Ser His Asn Ser Lys Phe Asp Phe				
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cac aag tac atc aag agg tcc atg gag gac gac ttc aaa gtc gtc gtt				946
His Lys Tyr Ile Lys Arg Ser Met Glu Asp Asp Phe Lys Val Val Val				
245		250	255	
ggc atc agc ctc ccg ctg tgg tgt gtg gcg atc ctc acc ctc ttc ctt				994
Gly Ile Ser Leu Pro Leu Trp Cys Val Ala Ile Leu Thr Leu Phe Leu				
260		265	270	
gac att gac ggg atc ggc acg ctc acc tgg att tct ttc atc cct ctc				1042
Asp Ile Asp Gly Ile Gly Thr Leu Thr Trp Ile Ser Phe Ile Pro Leu				
275		280	285	
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Val Ile Leu Leu Cys Val Gly Thr Lys Leu Glu Met Ile Ile Met Glu				
290		295	300	305
atg gcc ctg gag atc cag gac cgg gcg agc gtc atc aag ggg gcg ccc				1138
Met Ala Leu Glu Ile Gln Asp Arg Ala Ser Val Ile Lys Gly Ala Pro				
310		315	320	
gtg gtt gag ccc agc aac aag ttc ttc tgg ttc cac cgc ccc gac tgg				1186
Val Val Glu Pro Ser Asn Lys Phe Phe Trp Phe His Arg Pro Asp Trp				
325		330	335	
gtc ctc ttc ttc ata cac ctg acg cta ttc cag aac gcg ttt cag atg				1234
Val Leu Phe Phe Ile His Leu Thr Leu Phe Gln Asn Ala Phe Gln Met				
340		345	350	
gca cat ttc gtg tgg aca gtg gcc acg ccc ggc ttg aag aaa tgc ttc				1282

Ala His Phe Val Trp Thr Val Ala Thr Pro Gly Leu Lys Lys Cys Phe
 355 360 365

cat atg cac atc ggg ctg agc atc atg aag gtc gtg ctg ggg ctg gct 1330
 His Met His Ile Gly Leu Ser Ile Met Lys Val Val Leu Gly Leu Ala
 370 375 380 385

ctt cag ttc ctc tgc agc tat atc acc ttc ccg ctc tac gcg ctc gtc 1378
 Leu Gln Phe Leu Cys Ser Tyr Ile Thr Phe Pro Leu Tyr Ala Leu Val
 390 395 400

aca cag atg gga tca aac atg aag agg tcc atc ttc gac gag cag acg 1426
 Thr Gln Met Gly Ser Asn Met Lys Arg Ser Ile Phe Asp Glu Gln Thr
 405 410 415

gcc aag gcg ctg aca aac tgg cgg aac acg gcc aag gag aag aag aag 1474
 Ala Lys Ala Leu Thr Asn Trp Arg Asn Thr Ala Lys Glu Lys Lys Lys
 420 425 430

gtc cga gac acg gac atg ctg atg gcg cag atg atc ggc gac gcg acg 1522
 Val Arg Asp Thr Asp Met Leu Met Ala Gln Met Ile Gly Asp Ala Thr
 435 440 445

ccc agc cga ggg gcg tcg ccc atg cct agc cgg ggc tcg tcg cca gtg 1570
 Pro Ser Arg Gly Ala Ser Pro Met Pro Ser Arg Gly Ser Ser Pro Val
 450 455 460 465

cac ctg ctt cac aag ggc atg gga cgg tcc gac gat ccc cag agc acg 1618
 His Leu Leu His Lys Gly Met Gly Arg Ser Asp Asp Pro Gln Ser Thr
 470 475 480

cca acc tcg cca agg gcc atg gag gag gct agg gac atg tac ccg gtt 1666
 Pro Thr Ser Pro Arg Ala Met Glu Glu Ala Arg Asp Met Tyr Pro Val
 485 490 495

gtg gtg gcg cat cca gtg cac aga cta aat cct gct gac agg aga agg 1714
 Val Val Ala His Pro Val His Arg Leu Asn Pro Ala Asp Arg Arg Arg
 500 505 510

tcg gtc tcg tcg tcg gca ctc gat gtc gac att ccc agc gca gat ttt 1762
 Ser Val Ser Ser Ser Ala Leu Asp Val Asp Ile Pro Ser Ala Asp Phe
 515 520 525

tcc ttc agc cag gga tgagacaagt ttctgtattg atgtagtcc aatgtatagc 1817
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<212> PRT

<213> Triticum sp.

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 35 40 45
 Arg His Lys Asn Ala Leu Ala Glu Ala Leu Glu Lys Ile Lys Ala Glu
 50 55 60
 Leu Met Leu Val Gly Phe Ile Ser Leu Leu Leu Ala Val Thr Gln Asp
 65 70 75 80
 Pro Ile Ser Gly Ile Cys Ile Ser Glu Lys Ala Ala Ser Ile Met Arg
 85 90 95
 Pro Cys Lys Leu Pro Pro Gly Ser Val Lys Ser Lys Tyr Lys Asp Tyr
 100 105 110
 Tyr Cys Ala Lys Gln Gly Lys Val Ser Leu Met Ser Thr Gly Ser Leu
 115 120 125
 His Gln Leu His Ile Phe Ile Phe Val Leu Ala Val Phe His Val Thr
 130 135 140
 Tyr Ser Val Ile Ile Met Ala Leu Ser Arg Leu Lys Met Arg Thr Trp
 145 150 155 160
 Lys Lys Trp Glu Thr Glu Thr Ala Ser Leu Glu Tyr Gln Phe Ala Asn
 165 170 175
 Asp Pro Ala Arg Phe Arg Phe Thr His Gln Thr Ser Phe Val Lys Arg
 180 185 190
 His Leu Gly Leu Ser Ser Thr Pro Gly Val Arg Trp Val Val Ala Phe
 195 200 205
 Phe Arg Gln Phe Phe Arg Ser Val Thr Lys Val Asp Tyr Leu Thr Leu
 210 215 220
 Arg Ala Gly Phe Ile Asn Ala His Leu Ser His Asn Ser Lys Phe Asp
 225 230 235 240
 Phe His Lys Tyr Ile Lys Arg Ser Met Glu Asp Asp Phe Lys Val Val
 245 250 255
 Val Gly Ile Ser Leu Pro Leu Trp Cys Val Ala Ile Leu Thr Leu Phe
 260 265 270
 Leu Asp Ile Asp Gly Ile Gly Thr Leu Thr Trp Ile Ser Phe Ile Pro
 275 280 285

Leu Val Ile Leu Leu Cys Val Gly Thr Lys Leu Glu Met Ile Ile Met
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 Glu Met Ala Leu Glu Ile Gln Asp Arg Ala Ser Val Ile Lys Gly Ala
 305 310 315 320
 Pro Val Val Glu Pro Ser Asn Lys Phe Phe Trp Phe His Arg Pro Asp
 325 330 335
 Trp Val Leu Phe Phe Ile His Leu Thr Leu Phe Gln Asn Ala Phe Gln
 340 345 350
 Met Ala His Phe Val Trp Thr Val Ala Thr Pro Gly Leu Lys Lys Cys
 355 360 365
 Phe His Met His Ile Gly Leu Ser Ile Met Lys Val Val Leu Gly Leu
 370 375 380
 Ala Leu Gln Phe Leu Cys Ser Tyr Ile Thr Phe Pro Leu Tyr Ala Leu
 385 390 395 400
 Val Thr Gln Met Gly Ser Asn Met Lys Arg Ser Ile Phe Asp Glu Gln
 405 410 415
 Thr Ala Lys Ala Leu Thr Asn Trp Arg Asn Thr Ala Lys Glu Lys Lys
 420 425 430
 Lys Val Arg Asp Thr Asp Met Leu Met Ala Gln Met Ile Gly Asp Ala
 435 440 445
 Thr Pro Ser Arg Gly Ala Ser Pro Met Pro Ser Arg Gly Ser Ser Pro
 450 455 460
 Val His Leu Leu His Lys Gly Met Gly Arg Ser Asp Asp Pro Gln Ser
 465 470 475 480
 Thr Pro Thr Ser Pro Arg Ala Met Glu Glu Ala Arg Asp Met Tyr Pro
 485 490 495
 Val Val Val Ala His Pro Val His Arg Leu Asn Pro Ala Asp Arg Arg
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<223> location of the amino acid sequence set forth in
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Pro Ser Trp Ala Val Ala Leu Val Phe Ala Val Met Ile Ile Val Ser	
20 25 30	
gtc ctc ctg gag cac gcg ctc cac aag ctc ggc cat tgg ttc cac aag	144
Val Leu Leu Glu His Ala Leu His Lys Leu Gly His Trp Phe His Lys	
35 40 45	
cgg cac aag aac gcg ctg gcg gag gcg ctg gag aag atc aaa gcg gag	192
Arg His Lys Asn Ala Leu Ala Glu Ala Leu Glu Lys Ile Lys Ala Glu	
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ctg atg ctg gtg ggg ttc atc tcg ctg ctg ctc gcc gtg acg cag gac	240
Leu Met Leu Val Gly Phe Ile Ser Leu Leu Ala Val Thr Gln Asp	
65 70 75 80	
cca atc tcc ggg ata tgc atc tcc gag aag gcc gcc agc atc atg cgg	288
Pro Ile Ser Gly Ile Cys Ile Ser Glu Lys Ala Ala Ser Ile Met Arg	
85 90 95	
ccc tgc agc ctg ccc cct ggt tcc gtc aag agc aag tac aaa gac tac	336
Pro Cys Ser Leu Pro Pro Gly Ser Val Lys Ser Lys Tyr Lys Asp Tyr	
100 105 110	
tac tgc gcc aaa aag ggc aag gtg tcg cta atg tcc acg ggc agc ttg	384
Tyr Cys Ala Lys Lys Gly Lys Val Ser Leu Met Ser Thr Gly Ser Leu	
115 120 125	
cac cag ctc cac atg ttc atc ttc gtg ctc gcc gtc ttc cat gtc acc	432
His Gln Leu His Met Phe Ile Phe Val Leu Ala Val Phe His Val Thr	
130 135 140	
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Tyr Ser Val Ile Ile Met Ala Leu Ser Arg Leu Lys Met Arg Thr Trp	
145 150 155 160	

aag aaa tgg gag aca gag acc gyc tcc ttg gaa tac cag ttc gca aat	528
Lys Lys Trp Glu Thr Glu Thr Xaa Ser Leu Glu Tyr Gln Phe Ala Asn	
165 170 175	
gat cct gcg cgg ttc cgc ttc acg cac cag acg tcg ttc gtg aag cgt	576
Asp Pro Ala Arg Phe Arg Phe Thr His Gln Thr Ser Phe Val Lys Arg	
180 185 190	
cac ctg ggc ctc tcc agc acc ccc ggc atc aga tgg gtg gtg gcc ttc	624
His Leu Gly Leu Ser Ser Thr Pro Gly Ile Arg Trp Val Val Ala Phe	
195 200 205	
ttc agg cag ttc ttc agg tcg gtc acc aag gtg gac tac ctc acc ctg	672
Phe Arg Gln Phe Phe Arg Ser Val Thr Lys Val Asp Tyr Leu Thr Leu	
210 215 220	
agg gca ggc ttc atc aac gcg cat ttg tcg cat aac agc aag ttc gac	720
Arg Ala Gly Phe Ile Asn Ala His Leu Ser His Asn Ser Lys Phe Asp	
225 230 235 240	
ttc cac aag tac atc aag agg tcc atg gag gac gac ttc aaa gtc gtc	768
Phe His Lys Tyr Ile Lys Arg Ser Met Glu Asp Asp Phe Lys Val Val	
245 250 255	
gtt ggc atc agc ctc ccg ctg tgg tgt gtg gcg atc ctc acc ctc ttc	816
Val Gly Ile Ser Leu Pro Leu Trp Cys Val Ala Ile Leu Thr Leu Phe	
260 265 270	
ctt gat att gac ggg atc ggc acg ctc acc tgg att tct ttc atc cct	864
Leu Asp Ile Asp Gly Ile Gly Thr Leu Thr Trp Ile Ser Phe Ile Pro	
275 280 285	
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Leu Val Ile Leu Leu Cys Val Gly Thr Lys Leu Glu Met Ile Ile Met	
290 295 300	
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Glu Met Ala Leu Glu Ile Gln Asp Arg Ala Ser Val Ile Lys Gly Ala	
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Pro Val Val Glu Pro Ser Asn Lys Phe Phe Trp Phe His Arg Pro Asp	
325 330 335	
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Trp Val Leu Phe Phe Ile His Leu Thr Leu Phe Gln Asn Ala Phe Gln	
340 345 350	
atg gca cat ttc gtc tgg aca gtg gcc acg ccc ggc ttg aag aaa tgc	1104
Met Ala His Phe Val Trp Thr Val Ala Thr Pro Gly Leu Lys Lys Cys	
355 360 365	
ttc cat atg cac atc ggt ctg agc atc atg aag gtc gtg ctg ggg ctg	1152
Phe His Met His Ile Gly Leu Ser Ile Met Lys Val Val Leu Gly Leu	

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385	390	395	400
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Val Thr Gln Met Gly Ser Asn Met Lys Arg Ser Ile Phe Asp Glu Gln			
	405	410	415
acg gcc aag gcg ctg acc aac tgg cgg aac acg gcc aag gag aag aag			1296
Thr Ala Lys Ala Leu Thr Asn Trp Arg Asn Thr Ala Lys Glu Lys Lys			
	420	425	430
aag gtc cga gac acg gac atg ctg atg gcg cag atg atc ggc gac gcg			1344
Lys Val Arg Asp Thr Asp Met Leu Met Ala Gln Met Ile Gly Asp Ala			
	435	440	445
acg ccc agc cga ggc acg tgc ccg atg cct agc cgg gct tgc tca ccg			1392
Thr Pro Ser Arg Gly Thr Ser Pro Met Pro Ser Arg Ala Ser Ser Pro			
	450	455	460
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Val His Leu Leu His Lys Gly Met Gly Arg Ser Asp Asp Pro Gln Ser			
	465	470	475
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Ala Pro Thr Ser Pro Arg Thr Met Glu Glu Ala Arg Asp Met Tyr Pro			
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Val Val Val Ala His Pro Val His Arg Leu Asn Pro Ala Asp Arg Arg			
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Arg Ser Val Ser Ser Ser Ala Leu Asp Ala Asp Ile Pro Ser Ala Asp			
	515	520	525
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Phe Ser Phe Ser Gln Gly			
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 Arg His Lys Asn Ala Leu Ala Glu Ala Leu Glu Lys Ile Lys Ala Glu
 50 55 60
 Leu Met Leu Val Gly Phe Ile Ser Leu Leu Leu Ala Val Thr Gln Asp
 65 70 75 80
 Pro Ile Ser Gly Ile Cys Ile Ser Glu Lys Ala Ala Ser Ile Met Arg
 85 90 95
 Pro Cys Ser Leu Pro Pro Gly Ser Val Lys Ser Lys Tyr Lys Asp Tyr
 100 105 110
 Tyr Cys Ala Lys Lys Gly Lys Val Ser Leu Met Ser Thr Gly Ser Leu
 115 120 125
 His Gln Leu His Met Phe Ile Phe Val Leu Ala Val Phe His Val Thr
 130 135 140
 Tyr Ser Val Ile Ile Met Ala Leu Ser Arg Leu Lys Met Arg Thr Trp
 145 150 155 160
 Lys Lys Trp Glu Thr Glu Thr Xaa Ser Leu Glu Tyr Gln Phe Ala Asn
 165 170 175
 Asp Pro Ala Arg Phe Arg Phe Thr His Gln Thr Ser Phe Val Lys Arg
 180 185 190
 His Leu Gly Leu Ser Ser Thr Pro Gly Ile Arg Trp Val Val Ala Phe
 195 200 205
 Phe Arg Gln Phe Phe Arg Ser Val Thr Lys Val Asp Tyr Leu Thr Leu
 210 215 220
 Arg Ala Gly Phe Ile Asn Ala His Leu Ser His Asn Ser Lys Phe Asp
 225 230 235 240
 Phe His Lys Tyr Ile Lys Arg Ser Met Glu Asp Asp Phe Lys Val Val
 245 250 255
 Val Gly Ile Ser Leu Pro Leu Trp Cys Val Ala Ile Leu Thr Leu Phe
 260 265 270
 Leu Asp Ile Asp Gly Ile Gly Thr Leu Thr Trp Ile Ser Phe Ile Pro
 275 280 285
 Leu Val Ile Leu Leu Cys Val Gly Thr Lys Leu Glu Met Ile Ile Met
 290 295 300

Glu Met Ala Leu Glu Ile Gln Asp Arg Ala Ser Val Ile Lys Gly Ala
 305 310 315 320
 Pro Val Val Glu Pro Ser Asn Lys Phe Phe Trp Phe His Arg Pro Asp
 325 330 335
 Trp Val Leu Phe Phe Ile His Leu Thr Leu Phe Gln Asn Ala Phe Gln
 340 345 350
 Met Ala His Phe Val Trp Thr Val Ala Thr Pro Gly Leu Lys Lys Cys
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 Phe His Met His Ile Gly Leu Ser Ile Met Lys Val Val Leu Gly Leu
 370 375 380
 Ala Leu Gln Phe Leu Cys Ser Tyr Ile Thr Phe Pro Leu Tyr Ala Leu
 385 390 395 400
 Val Thr Gln Met Gly Ser Asn Met Lys Arg Ser Ile Phe Asp Glu Gln
 405 410 415
 Thr Ala Lys Ala Leu Thr Asn Trp Arg Asn Thr Ala Lys Glu Lys Lys
 420 425 430
 Lys Val Arg Asp Thr Asp Met Leu Met Ala Gln Met Ile Gly Asp Ala
 435 440 445
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 450 455 460
 Val His Leu Leu His Lys Gly Met Gly Arg Ser Asp Asp Pro Gln Ser
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 Val Val Val Ala His Pro Val His Arg Leu Asn Pro Ala Asp Arg Arg
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Lys Met Arg Thr Trp Lys Lys Trp Glu Thr Glu Thr Ala Ser Leu Glu	
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Tyr Gln Phe Ala Asn Asp Pro Ala Arg Phe Arg Phe Thr His Gln Thr	
175 180 185	
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Ser Phe Val Lys Arg His Leu Gly Leu Ser Ser Thr Pro Gly Val Arg	
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Trp Val Val Ala Phe Phe Arg Gln Phe Phe Arg Ser Val Thr Lys Val	
205 210 215	
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Asp Tyr Leu Thr Leu Arg Ala Gly Phe Ile Asn Ala His Leu Ser Gln	
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Asn Ser Lys Phe Asp Phe His Lys Tyr Ile Lys Arg Ser Met Glu Asp	
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Asp Phe Lys Val Val Val Gly Ile Ser Leu Pro Leu Trp Ala Val Ala	
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Ile Leu Thr Leu Phe Leu Asp Ile Asp Gly Ile Gly Thr Leu Thr Trp	
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Val Ser Phe Ile Pro Leu Ile Ile Leu Leu Cys Val Gly Thr Lys Leu	
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Glu Met Ile Ile Met Gly Met Ala Leu Glu Ile Gln Asp Arg Ser Ser	
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Gln Asn Ala Phe Gln Met Ala His Phe Val Trp Thr Val Ala Thr Pro	
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 Arg Asp Met Tyr Pro Val Val Val Ala His Pro Val His Arg Leu Asn
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 Pro Ala Asp Arg Arg Arg Ser Val Ser Ser Ser Ala Leu Asp Ala Asp
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 Ile Pro Ser Ala Asp Phe Ser Phe Ser Gln Gly
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<212> PRT

<213> Triticum sp.

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 35 40 45
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 Leu Met Leu Val Gly Phe Ile Ser Leu Leu Leu Ala Val Thr Gln Asp
 65 70 75 80
 Pro Ile Ser Gly Ile Cys Ile Ser Gln Lys Ala Ala Ser Ile Met Arg
 85 90 95
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 100 105 110
 Tyr Cys Ala Lys Glu Gly Lys Val Ala Leu Met Ser Thr Gly Ser Leu
 115 120 125
 His Gln Leu His Ile Phe Ile Phe Val Leu Ala Val Phe His Val Thr
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 Tyr Ser Val Ile Ile Met Ala Leu Ser Arg Leu Lys Met Arg Thr Trp
 145 150 155 160
 Lys Lys Trp Glu Thr Glu Thr Ala Ser Leu Glu Tyr Gln Phe Ala Asn
 165 170 175
 Asp Pro Ala Arg Phe Arg Phe Thr His Gln Thr Ser Phe Val Lys Arg
 180 185 190
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 195 200 205
 Phe Arg Gln Phe Phe Arg Ser Val Thr Lys Val Asp Tyr Leu Thr Leu
 210 215 220
 Arg Ala Gly Phe Ile Asn Ala His Leu Ser Gln Asn Ser Lys Phe Asp
 225 230 235 240
 Phe His Lys Tyr Ile Lys Arg Ser Met Glu Asp Asp Phe Lys Val Val
 245 250 255
 Val Gly Ile Ser Leu Pro Leu Trp Ala Val Ala Ile Leu Thr Leu Phe
 260 265 270
 Leu Asp Ile Asp Gly Ile Gly Thr Leu Thr Trp Val Ser Phe Ile Pro
 275 280 285

Leu Ile Ile Leu Leu Cys Val Gly Thr Lys Leu Glu Met Ile Ile Met
 290 295 300
 Gly Met Ala Leu Glu Ile Gln Asp Arg Ser Ser Val Ile Lys Gly Ala
 305 310 315 320
 Pro Val Val Glu Pro Ser Asn Lys Phe Phe Trp Phe His Arg Pro Asp
 325 330 335
 Trp Val Leu Phe Phe Ile His Leu Thr Leu Phe Gln Asn Ala Phe Gln
 340 345 350
 Met Ala His Phe Val Trp Thr Val Ala Thr Pro Gly Leu Lys Asp Cys
 355 360 365
 Phe His Met Asn Ile Gly Leu Ser Ile Met Lys Val Val Leu Gly Leu
 370 375 380
 Ala Leu Gln Phe Leu Cys Ser Tyr Ile Thr Phe Pro Leu Tyr Ala Leu
 385 390 395 400
 Val Thr Gln Met Gly Ser Asn Met Lys Arg Ser Ile Phe Asp Glu Gln
 405 410 415
 Thr Ala Lys Ala Leu Thr Asn Trp Arg Asn Thr Ala Lys Glu Lys Lys
 420 425 430
 Lys Val Arg Asp Thr Asp Met Leu Met Ala Gln Met Ile Gly Asp Ala
 435 440 445
 Thr Pro Ser Arg Gly Thr Ser Pro Met Pro Ser Arg Gly Ser Ser Pro
 450 455 460
 Val His Leu Leu Gln Lys Gly Met Gly Arg Ser Asp Asp Pro Gln Ser
 465 470 475 480
 Ala Pro Thr Ser Pro Arg Thr Met Glu Glu Ala Arg Asp Met Tyr Pro
 485 490 495
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Val Ala Thr Trp Leu Trp Glu Lys His Lys Asn Ser Leu Leu Glu Ala
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Leu Leu Thr Phe Gly Glu Gln Tyr Ile Leu Lys Ile Cys Ile Pro Glu
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Lys Ala Ala Ala Ser Met Leu Pro Cys Pro Ala Pro Ser Thr His Asp
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- 22 -

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Asn	Val	Asn	Gly	Trp	His	Thr	Tyr	Phe	Trp	Ile	Ala	Phe	Ile	Pro	Phe		
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gct	ttg	ctt	ctt	gct	gtg	gga	aca	aag	ttg	gag	cat	gtg	att	gca	cag	1066	
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Leu	Ala	His	Glu	Val	Ala	Glu	Lys	His	Val	Ala	Ile	Glu	Gly	Asp	Leu		
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Val	Val	Lys	Pro	Ser	Asp	Glu	His	Phe	Trp	Phe	Ser	Lys	Pro	Gln	Ile		
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Val	Leu	Tyr	Leu	Ile	His	Phe	Ile	Leu	Phe	Gln	Asn	Ala	Phe	Glu	Ile		
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Ala	Phe	Phe	Phe	Trp	Ile	Trp	Val	Thr	Tyr	Gly	Phe	Asp	Ser	Cys	Ile		
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Ser Gln Met Gly Ser Ser Phe Lys Lys Ala Ile Phe Glu Glu Asn Val
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cag gtt ggt ctt gtt ggt tgg gca cag aaa gtg aaa caa aag aga gac 1450
Gln Val Gly Leu Val Gly Trp Ala Gln Lys Val Lys Gln Lys Arg Asp
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Leu Lys Ala Ala Ala Ser Asn Gly Asn Glu Gly Ser Ser Gln Ala Gly
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Pro Gly Pro Asp Ser Gly Ser Gly Ser Ala Pro Ala Ala Gly Pro Gly
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gca ggt ttt gca gga att cag ctc agc aga gta aca aga aac aac gca 1594
Ala Gly Phe Ala Gly Ile Gln Leu Ser Arg Val Thr Arg Asn Asn Ala
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ggg gac aca aac aat gag att aca cct gat cat aac aac tgagcagaga 1643
Gly Asp Thr Asn Asn Glu Ile Thr Pro Asp His Asn Asn
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Lys Pro Leu Tyr Glu Ala Leu Gln Lys Val Lys Glu Glu Leu Met Leu
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Leu Gly Phe Ile Ser Leu Leu Leu Thr Val Phe Gln Gly Leu Ile Ser
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 Ile Phe Ile Phe Val Leu Ala Ile Ser His Val Thr Phe Cys Val Leu
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 Asp Ser Ile Ala Asp Glu Lys Phe Asp Pro Glu Thr Ala Leu Arg Lys
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 Arg Arg Val Thr His Val His Asn His Ala Phe Ile Lys Glu His Phe
 210 215 220
 Leu Gly Ile Gly Lys Asp Ser Val Ile Leu Gly Trp Thr Gln Ser Phe
 225 230 235 240
 Leu Lys Gln Phe Tyr Asp Ser Val Thr Lys Ser Asp Tyr Val Thr Leu
 245 250 255
 Arg Leu Gly Phe Ile Met Thr His Cys Lys Gly Asn Pro Lys Leu Asn
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 Phe His Lys Tyr Met Met Arg Ala Xaa Glu Asp Asp Phe Lys Gln Val
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 305 310 315 320
 Phe Ala Leu Leu Leu Ala Val Gly Thr Lys Leu Glu His Val Ile Ala
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 Gln Leu Ala His Glu Val Ala Glu Lys His Val Ala Ile Glu Gly Asp
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 Leu Val Val Lys Pro Ser Asp Glu His Phe Trp Phe Ser Lys Pro Gln
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- 32 -

Ile Val Leu Tyr Leu Ile His Phe Ile Leu Phe Gln Asn Ala Phe Glu
 370 375 380
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 405 410 415
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Val Ala Ala Val Cys Thr Phe Phe Ile Val Val Ser Val Leu Leu Glu				
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Lys Leu Leu His Lys Val Gly Lys Val Leu Trp Asp Arg His Lys Thr				
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Gly Phe Ile Ser Leu Leu Leu Thr Phe Gly Gln Thr Tyr Ile Leu Asp				
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Ile Cys Ile Pro Ser His Val Ala Arg Thr Met Leu Pro Cys Pro Ala				
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 20 25 30
 gtt gaa cgc gcc att tac aga ttt gga aag tgg tta aag aag act aga 144
 Val Glu Arg Ala Ile Tyr Arg Phe Gly Lys Trp Leu Lys Lys Thr Arg
 35 40 45
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 Arg Lys Ala Leu Phe Thr Ser Leu Glu Lys Met Lys Glu Glu Leu Met
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 65 70 75 80
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 85 90 95
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 Cys Ser Glu Glu Asp Tyr Gly Ile His Lys Lys Val Leu Leu Glu His
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 115 120 125
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 130 135 140
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Val	Leu	Tyr	Ser	Gly	Ile	Ala	Ile	Gly	Leu	Ala	Met	Ser	Lys	Ile	Tyr	
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Ser	Trp	Arg	Lys	Trp	Glu	Ala	Gln	Ala	Ile	Ile	Met	Ala	Glu	Ser	Asp	
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		355					360					365				

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att tct ggg gtt ctc gtt cag ttc tgg tgt agt tat ggc act gtg cct 1152
 Ile Ser Gly Val Leu Val Gln Phe Trp Cys Ser Tyr Gly Thr Val Pro
 370 375 380

ctc aat gta atc gtt act cag atg gga tct cgg cat aag aaa gct gtg 1200
 Leu Asn Val Ile Val Thr Gln Met Gly Ser Arg His Lys Lys Ala Val
 385 390 395 400

ata gca gag agc gta aga gac tca ctt cac agt tgg tgc aag aga gtg 1248
 Ile Ala Glu Ser Val Arg Asp Ser Leu His Ser Trp Cys Lys Arg Val
 405 410 415

aaa gag agg tct aag cac acg aga tca gtg tgt tcc ctt gac aca gca 1296
 Lys Glu Arg Ser Lys His Thr Arg Ser Val Cys Ser Leu Asp Thr Ala
 420 425 430

aca ata gac gag aga gac gag atg aca gtg ggg aca ttg tct agg agc 1344
 Thr Ile Asp Glu Arg Asp Glu Met Thr Val Gly Thr Leu Ser Arg Ser
 435 440 445

tca tcg atg act tca ctg aat cag att acc ata aac tcc ata gac caa 1392
 Ser Ser Met Thr Ser Leu Asn Gln Ile Thr Ile Asn Ser Ile Asp Gln
 450 455 460

gca gag tct ata ttc gga gca gca gct tca tcc agc agt cct caa gat 1440
 Ala Glu Ser Ile Phe Gly Ala Ala Ala Ser Ser Ser Ser Pro Gln Asp
 465 470 475 480

gga tac acg tcg agg gtg gaa gaa tat ctg tct gaa aca tac aat aac 1488
 Gly Tyr Thr Ser Arg Val Glu Glu Tyr Leu Ser Glu Thr Tyr Asn Asn
 485 490 495

atc ggt tcg ata ccg cct tta aac gat gag att gag att gag att gaa 1536
 Ile Gly Ser Ile Pro Pro Leu Asn Asp Glu Ile Glu Ile Glu Ile Glu
 500 505 510

ggt gaa gaa gat aat gga ggg aga gga agt ggg agt gat gag aat aac 1584
 Gly Glu Glu Asp Asn Gly Gly Arg Gly Ser Gly Ser Asp Glu Asn Asn
 515 520 525

ggt gat gct gga gaa aca ctt ctt gag ttg ttt agg agg act tga 1629
 Gly Asp Ala Gly Glu Thr Leu Leu Glu Leu Phe Arg Arg Thr
 530 535 540

<210> 18

<211> 542

<212> PRT

<213> Arabidopsis thaliana

<400> 18

Met Glu His Met Met Lys Glu Gly Arg Ser Leu Ala Glu Thr Pro Thr
 1 5 10 15

Tyr Ser Val Ala Ser Val Val Thr Val Leu Val Phe Val Cys Phe Leu

Val	Glu	Arg	Ala	Ile	Tyr	Arg	Phe	Gly	Lys	Trp	Leu	Lys	Lys	Thr	Arg		
			35													40	45
Arg	Lys	Ala	Leu	Phe	Thr	Ser	Leu	Glu	Lys	Met	Lys	Glu	Glu	Leu	Met		
			50				55					60					
Leu	Leu	Gly	Leu	Ile	Ser	Leu	Leu	Leu	Ser	Gln	Ser	Ala	Arg	Trp	Ile		
			65			70									80		
Ser	Glu	Ile	Cys	Val	Asn	Ser	Ser	Leu	Phe	Asn	Ser	Lys	Phe	Tyr	Ile		
				85						90					95		
Cys	Ser	Glu	Glu	Asp	Tyr	Gly	Ile	His	Lys	Lys	Val	Leu	Leu	Glu	His		
			100					105						110			
Thr	Ser	Ser	Thr	Asn	Gln	Ser	Ser	Leu	Pro	His	His	Gly	Ile	His	Glu		
			115				120						125				
Ala	Ser	His	Gln	Cys	Gly	His	Gly	Arg	Glu	Pro	Phe	Val	Ser	Tyr	Glu		
			130				135					140					
Gly	Leu	Glu	Gln	Leu	Leu	Arg	Phe	Leu	Phe	Val	Leu	Gly	Ile	Thr	His		
			145			150				155					160		
Val	Leu	Tyr	Ser	Gly	Ile	Ala	Ile	Gly	Leu	Ala	Met	Ser	Lys	Ile	Tyr		
				165					170						175		
Ser	Trp	Arg	Lys	Trp	Glu	Ala	Gln	Ala	Ile	Ile	Met	Ala	Glu	Ser	Asp		
			180					185					190				
Ile	His	Leu	Cys	Phe	Leu	Arg	Gln	Phe	Arg	Gly	Ser	Ile	Arg	Lys	Ser		
			195				200					205					
Asp	Tyr	Phe	Ala	Leu	Arg	Leu	Gly	Phe	Leu	Thr	Lys	His	Asn	Leu	Pro		
			210			215						220					
Phe	Thr	Tyr	Asn	Phe	His	Met	Tyr	Met	Val	Arg	Thr	Met	Glu	Asp	Glu		
			225			230					235				240		
Phe	His	Gly	Ile	Val	Gly	Ile	Ser	Trp	Pro	Leu	Trp	Val	Tyr	Ala	Ile		
				245						250					255		
Val	Cys	Ile	Cys	Ile	Asn	Val	His	Gly	Leu	Asn	Met	Tyr	Phe	Trp	Ile		
			260					265						270			
Ser	Phe	Val	Pro	Ala	Ile	Leu	Val	Met	Leu	Val	Gly	Thr	Lys	Leu	Glu		
			275				280					285					
His	Val	Val	Ser	Lys	Leu	Ala	Leu	Glu	Val	Lys	Glu	Gln	Gln	Thr	Gly		
			290			295					300						
Thr	Ser	Asn	Gly	Ala	Gln	Val	Lys	Pro	Arg	Asp	Gly	Leu	Phe	Trp	Phe		
					310					315					320		

- 42 -

Gly Lys Pro Glu Ile Leu Leu Arg Leu Ile Gln Phe Ile Ile Phe Gln
 325 330 335
 Asn Ala Phe Glu Met Ala Thr Phe Ile Trp Phe Leu Trp Gly Ile Lys
 340 345 350
 Glu Arg Ser Cys Phe Met Lys Asn His Val Met Ile Ser Ser Arg Leu
 355 360 365
 Ile Ser Gly Val Leu Val Gln Phe Trp Cys Ser Tyr Gly Thr Val Pro
 370 375 380
 Leu Asn Val Ile Val Thr Gln Met Gly Ser Arg His Lys Lys Ala Val
 385 390 395 400
 Ile Ala Glu Ser Val Arg Asp Ser Leu His Ser Trp Cys Lys Arg Val
 405 410 415
 Lys Glu Arg Ser Lys His Thr Arg Ser Val Cys Ser Leu Asp Thr Ala
 420 425 430
 Thr Ile Asp Glu Arg Asp Glu Met Thr Val Gly Thr Leu Ser Arg Ser
 435 440 445
 Ser Ser Met Thr Ser Leu Asn Gln Ile Thr Ile Asn Ser Ile Asp Gln
 450 455 460
 Ala Glu Ser Ile Phe Gly Ala Ala Ala Ser Ser Ser Ser Pro Gln Asp
 465 470 475 480
 Gly Tyr Thr Ser Arg Val Glu Glu Tyr Leu Ser Glu Thr Tyr Asn Asn
 485 490 495
 Ile Gly Ser Ile Pro Pro Leu Asn Asp Glu Ile Glu Ile Glu Ile Glu
 500 505 510
 Gly Glu Glu Asp Asn Gly Gly Arg Gly Ser Gly Ser Asp Glu Asn Asn
 515 520 525
 Gly Asp Ala Gly Glu Thr Leu Leu Glu Leu Phe Arg Arg Thr
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<210> 19

<211> 20

<212> DNA

<213> Artificial Sequence

<220>

 <223> Description of Artificial Sequence:
 Oligonucleotide

<400> 19

atgtcggaca aaaaaggggt

<210> 20
<211> 20
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide

<400> 20
atgctaccac acgcagatcg

20

<210> 21
<211> 21
<212> DNA
<213> Artificial Sequence

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<400> 21
acagagacca cctccttgga a

21

<210> 22
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<400> 22
cagaaacttg tctcatccct gg

22

<210> 23
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<212> DNA
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<400> 23
aagaactgcc tgaagaaggc

20

<210> 24

<211> 20
<212> DNA
<213> Artificial Sequence

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20

<210> 25
<211> 20
<212> DNA
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<400> 25
ttccagcacc ggcacaagaa

20

<210> 26
<211> 28
<212> DNA
<213> Artificial Sequence

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Oligonucleotide

<400> 26
tggacctctt catgttcgat cccatctg

28

<210> 27
<211> 27
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide

<400> 27
cctgacgctg ttccagaatg cgtttca

27

<210> 28
<211> 20
<212> DNA
<213> Artificial Sequence

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<223> Description of Artificial Sequence:
Oligonucleotide

<400> 28

acttctgcag gtcgactcta

20

<210> 29

<211> 30

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:
Oligonucleotide

<400> 29

aagatcaaga tgaggacgtg gaagtcgtgg

30

<210> 30

<211> 30

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:
Oligonucleotide

<400> 30

aggctgaacc actggggcgc ctctcaccac

30

<210> 31

<211> 30

<212> DNA

<213> Artificial Sequence

<220>

<223> Description of Artificial Sequence:
Oligonucleotide

<400> 31

caagtatatg atgcgcgctc tagaggatga

30

<210> 32

<211> 30

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:

Oligonucleotide

<400> 32
aggtttcacc actaagtctc cttcaatggc 30

<210> 33
<211> 30
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide

<400> 33
gatcattcaa gacttaggct cactcatggag 30

<210> 34
<211> 30
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide

<400> 34
aacagcaagg aagattacaa atgatgccca 30

<210> 35
<211> 18
<212> DNA
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<220>
<223> Description of Artificial Sequence:
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<400> 35
ggattaagat ctaatggc 18

<210> 36
<211> 23
<212> DNA
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<220>
<223> Description of Artificial Sequence:
Oligonucleotide

<400> 36

caaagatctt catttcttaa aag

23

<210> 37

<211> 25

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:
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<400> 37

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25

<210> 38

<211> 25

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:
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<400> 38

gcggatcctc atccctggct gaagg

25

<210> 39

<211> 23

<212> DNA

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<223> Description of Artificial Sequence:
Oligonucleotide

<400> 39

ggatccacca tggccacaag atg

23

<210> 40

<211> 24

<212> DNA

<213> Artificial Sequence

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<223> Description of Artificial Sequence:
Oligonucleotide

<400> 40

ggatccttag tcaatatcat tagc

24

<210> 41
<211> 27
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide

<400> 41
gcggatccat gggtcacgga ggagaag

27

<210> 42
<211> 27
<212> DNA
<213> Artificial Sequence

<220>
<223> Description of Artificial Sequence:
Oligonucleotide

<400> 42
gcggatcctc agttggtatg atcagga

27